

Bone marrow adipose tissue influences response to treatment and metabolism of multiple myeloma cells

Chaimae Nouasria^{1,2}, Dávid Ernst^{1,2}, Béla Kajtár³, Zoltán Kohl⁴, Hussain Alizadeh⁴, Zoltán Patonai⁵, Balázs Radnai⁶, Eszter Vámos⁶, Zoltán Kellermayer^{1,2}

- ¹University of Pécs Clinical Center, Department of Immunology and Biotechnology
²University of Pécs Szentágotthai Research Centre, Multiple Myeloma Microenvironment Research Group
³University of Pécs Clinical Center, Department of Pathology
⁴University of Pécs Clinical Center, 1st Department of Internal Medicine, Division of Hematology
⁵University of Pécs Clinical Center, Department of Traumatology and Hand Surgery
⁶University of Pécs Medical school, Department of Biochemistry



Introduction

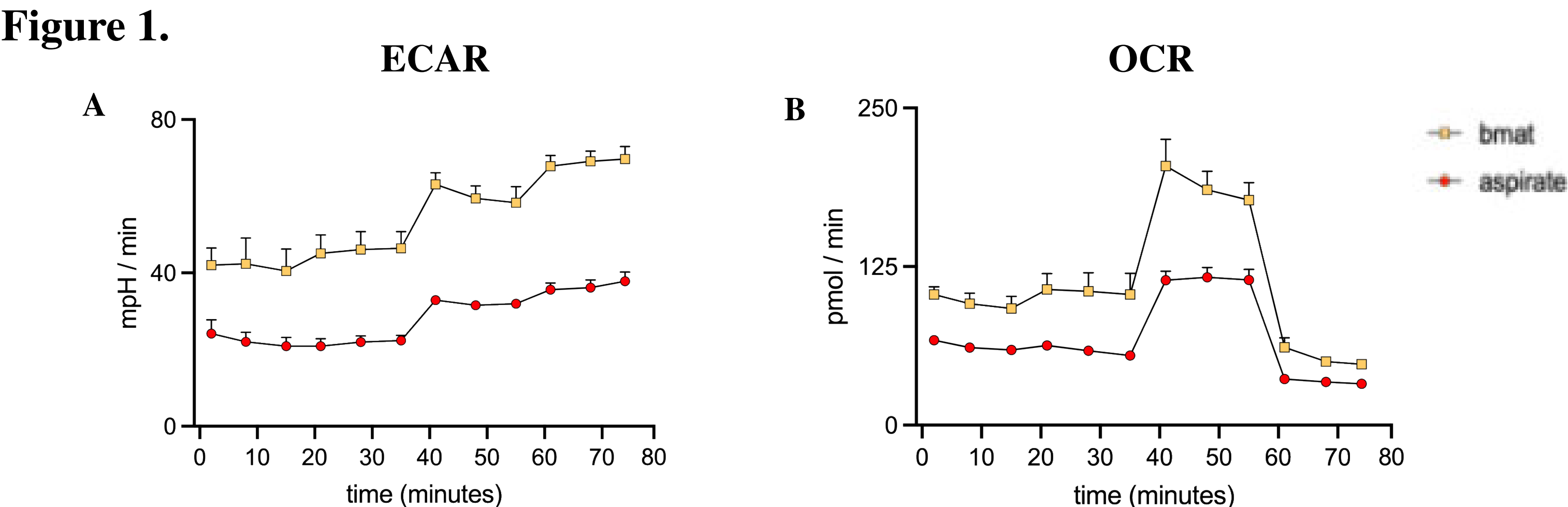
Multiple myeloma (MM) is a hematologic malignancy characterized by the clonal expansion of malignant plasma cells within the bone marrow (BM). The disease progression is largely influenced by the bone marrow microenvironment (BMM). Bone marrow adipocytes (BMAs) are one of the most abundant cells in the BM, suggesting they might play a role in disease course. We have recently described that bone marrow adipose tissue (BMAT) contains a higher percentage of MM cells. Our aim was to investigate whether these “fat-associated” MM cells differ from the “aspirate-associated” MM.

Materials and Methods

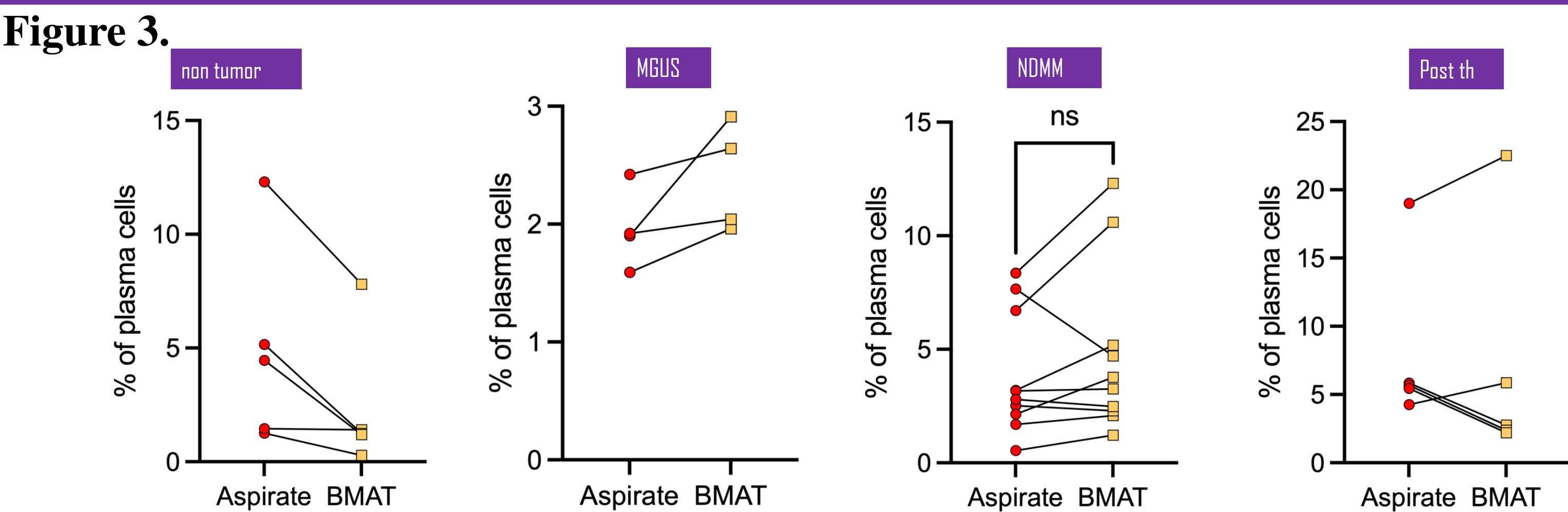
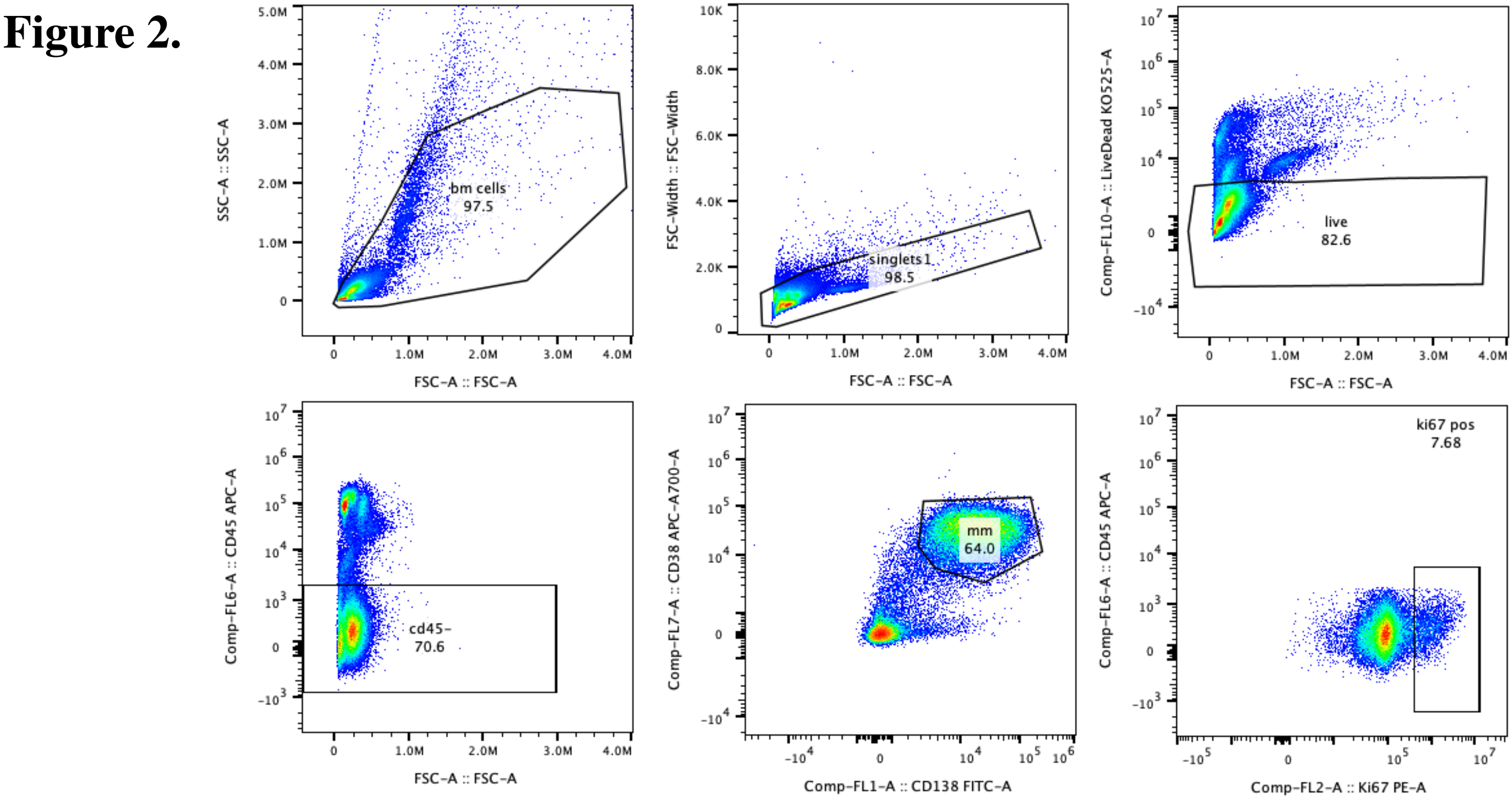
We isolated BMAT from aspirates of precursor stage (MGUS), newly diagnosed and follow-up MM patients and non-tumor controls. This BMAT fraction was then enzymatically digested to create a single-cell suspension, followed by red blood cell lysis. We then **A)** measured metabolism; **B)** compared proliferation rates of fat-associated vs aspirate-associated plasma cells; and **C)** compared the responses to conventional anti-MM drugs (bortezomib, thalidomide, melphalan, dexamethasone).

Results

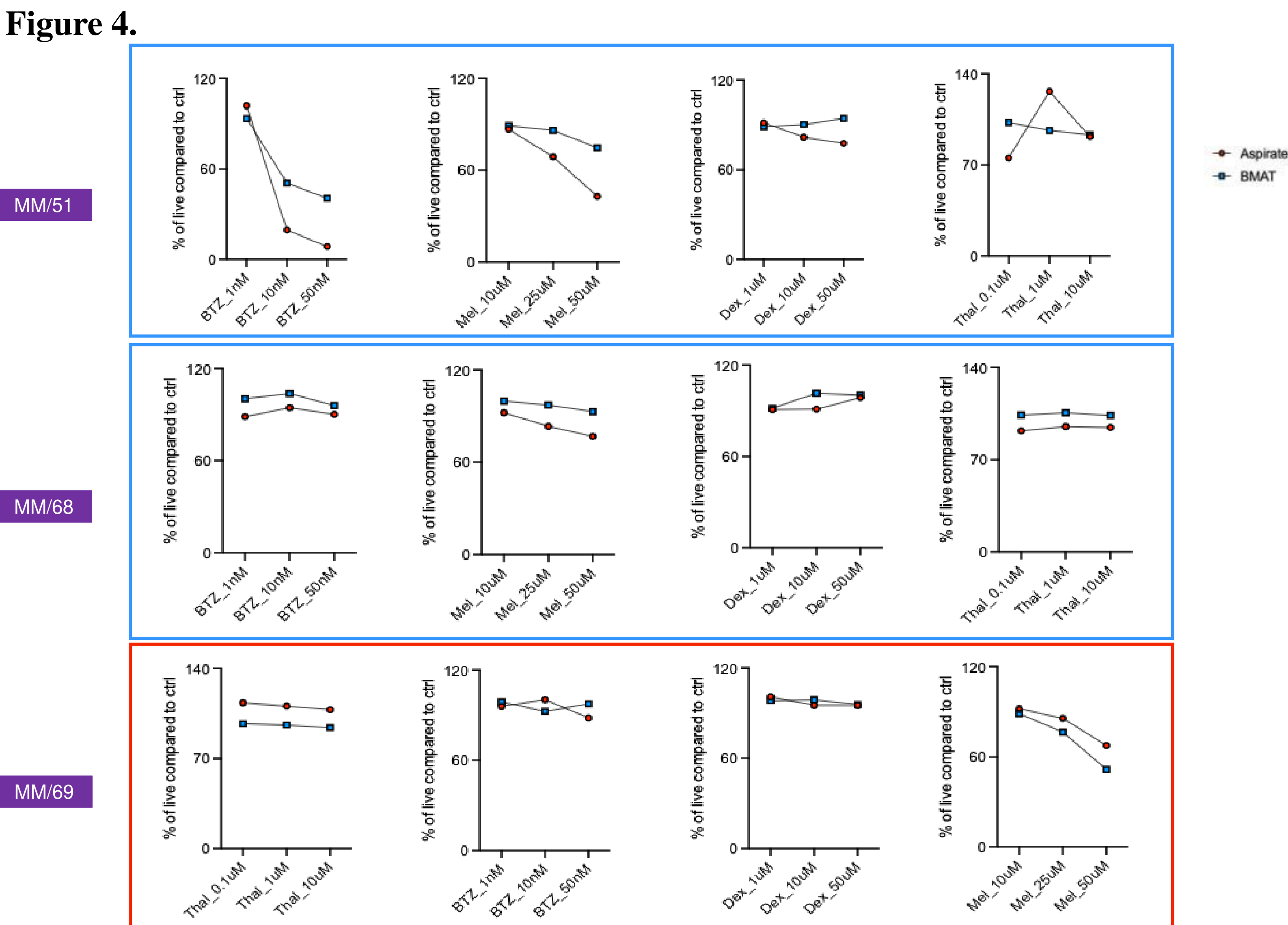
We performed Seahorse assays to assess the metabolic profile of the two fractions. We found that cells derived from the BMAT exhibit a higher glycolysis (**Fig1A**) and oxidative phosphorylation (**Fig1B**) compared to the cells from the aspirate, indicating enhanced metabolic activity of cells in the vicinity of BMAs.



We also performed Ki67 flow cytometric staining (**Fig2**), which revealed a lower percentage of proliferating fat-associated plasma cells in non-tumor controls but a higher percentage in MGUS. In NDMM and follow-up samples we observed mixed results (**Fig3**).



After the 4 hours treatment we found that in a subset of patients BMAT-associated MM cells responded less to drugs compared to aspirate-associated MM (**MM/58** and **MM/51**, **Fig4**). Meanwhile, in one patient it had the opposite effect (**MM/69**, **Fig4**).



Conclusions

Our data suggests that the BMAT might act as a supportive niche for MM by influencing MM cell metabolism, proliferation, and response to therapy. We plan to perform transcriptomic studies and *in vivo* mouse experiments to identify the exact molecular interactions between BMAs and MM cells.

Funding

Contact information

nouasria.chaimae@pte.hu
kellermayer.zoltan@pte.hu

SUPPORTED BY THE ÚNKP-23-5 NEW NATIONAL EXCELLENCE PROGRAM OF THE MINISTRY FOR CULTURE AND INNOVATION FROM THE SOURCE OF THE NATIONAL RESEARCH, DEVELOPMENT AND INNOVATION FUND.
This project was supported by the jános bolyai research scholarship of the hungarian academy of sciences
PTE ÁOK KA-2023-28
OTKA FK 146541



PROGRAM FINANCED
FROM THE NRDI FUND