

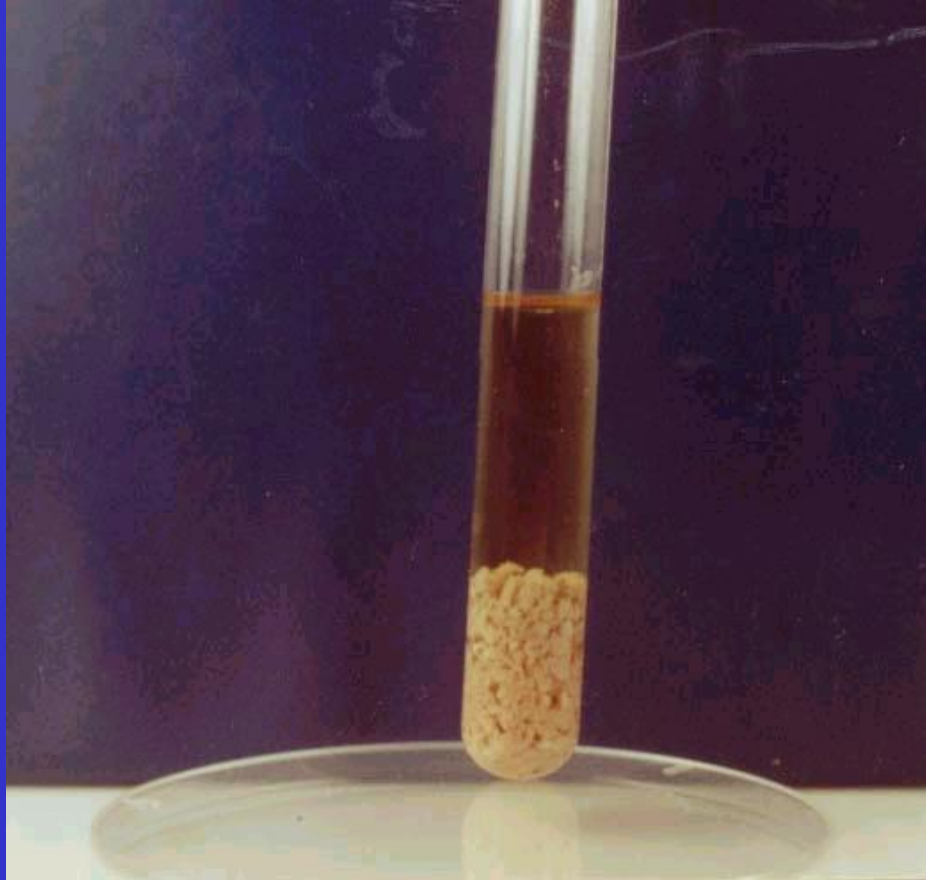
Practical examination

I. Sterile media

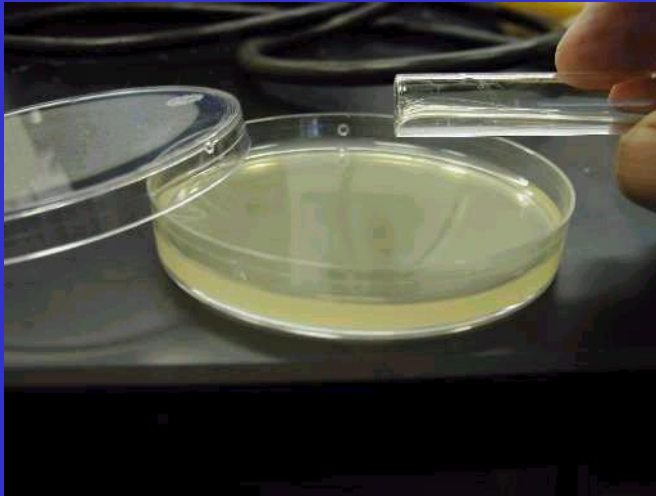
1. Bouillon, 2. Slant agar, tube agar



4. Enrichment media: meat bouillon



3., 5., 6.: Agar, blood agar and chocolate agar plates



7. Selective and differentiating media

c. Eosin methylene blue (EMB)

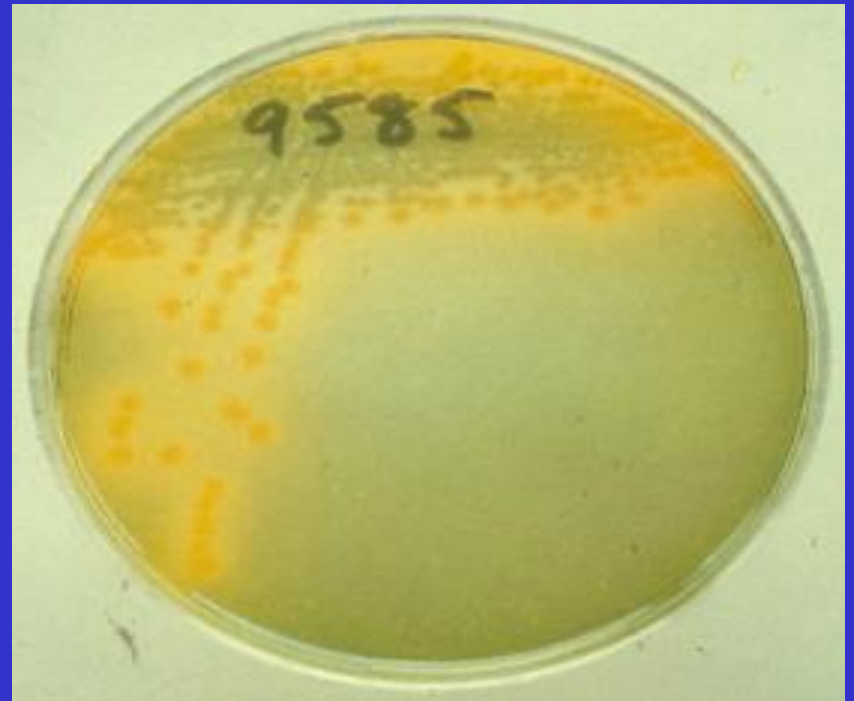
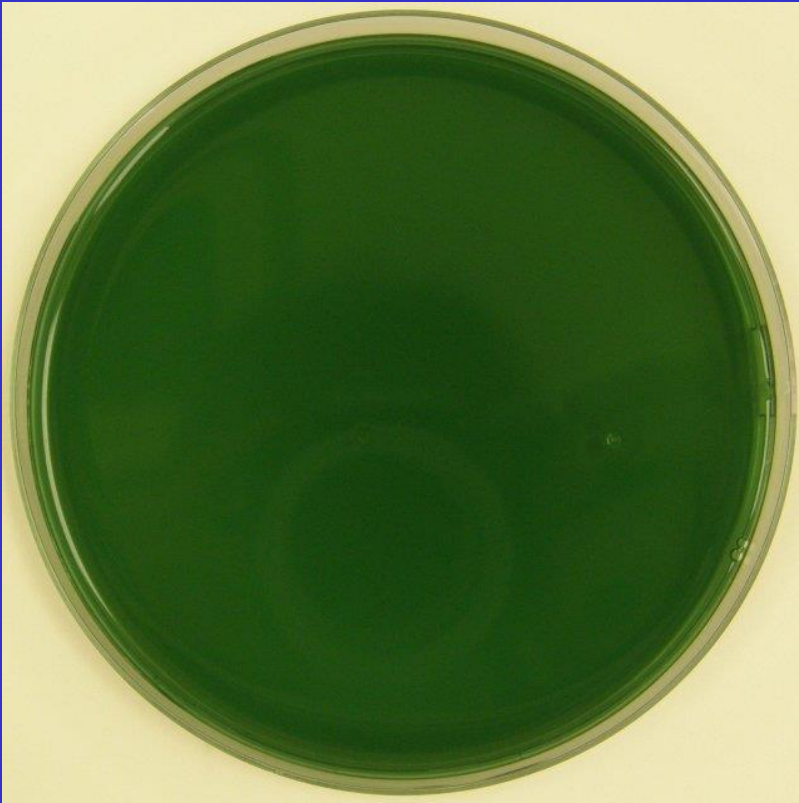


Lactose + colonies



Lactose - colonies

7. Selective and differentiating media g. TCBS



Vibrio cholerae

8. Transport media

Transport swabs
(e.g. Stuart)



9. Blood culture



10. Virus transport medium



Virus Transport Medium - Store at 4°C

NAME

WARD or DR.DATE.....

SITE

Exp:08/01

Break / cut swab into medium - Dispatch to:

Canterbury Health Laboratories

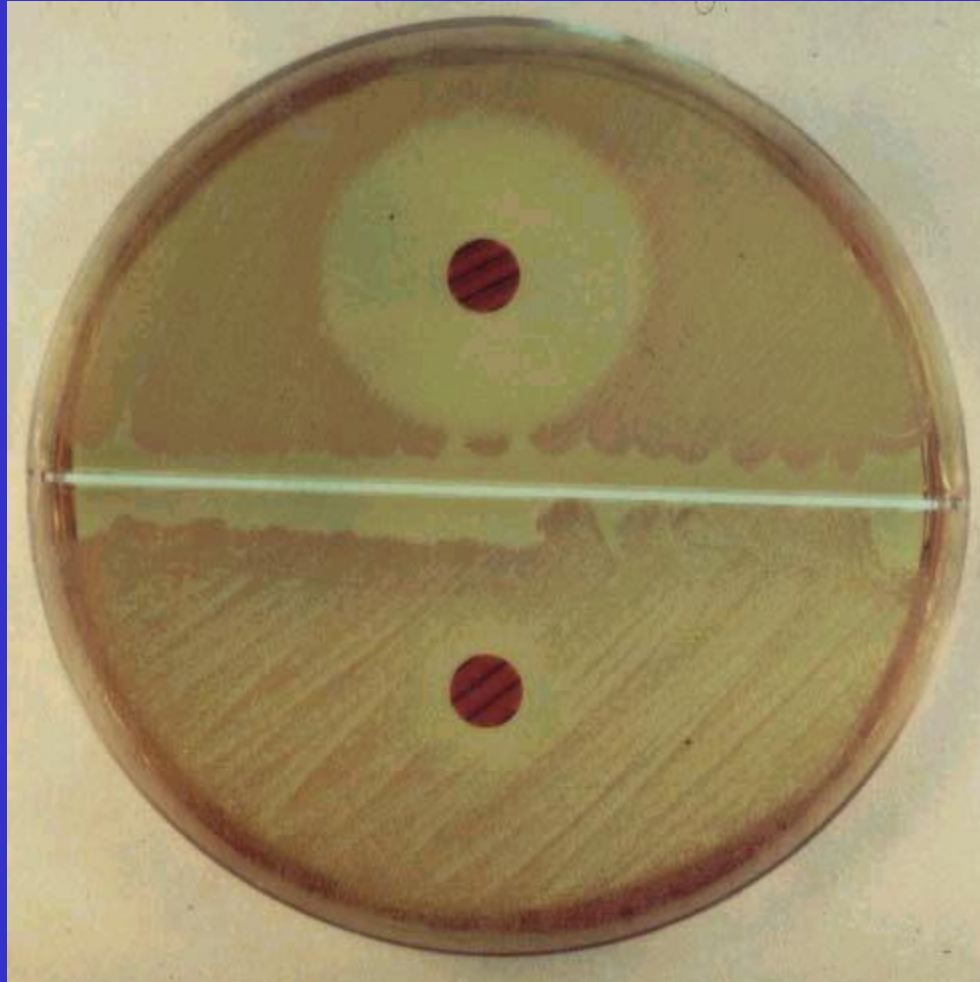


II. Cultures

1-2-3. *Staphylococcus aureus* and *S. epidermidis* on agar plate and blood agar plate



4. *S. epidermidis* (S) and *S. saprophyticus* (R) with novobiocin disc

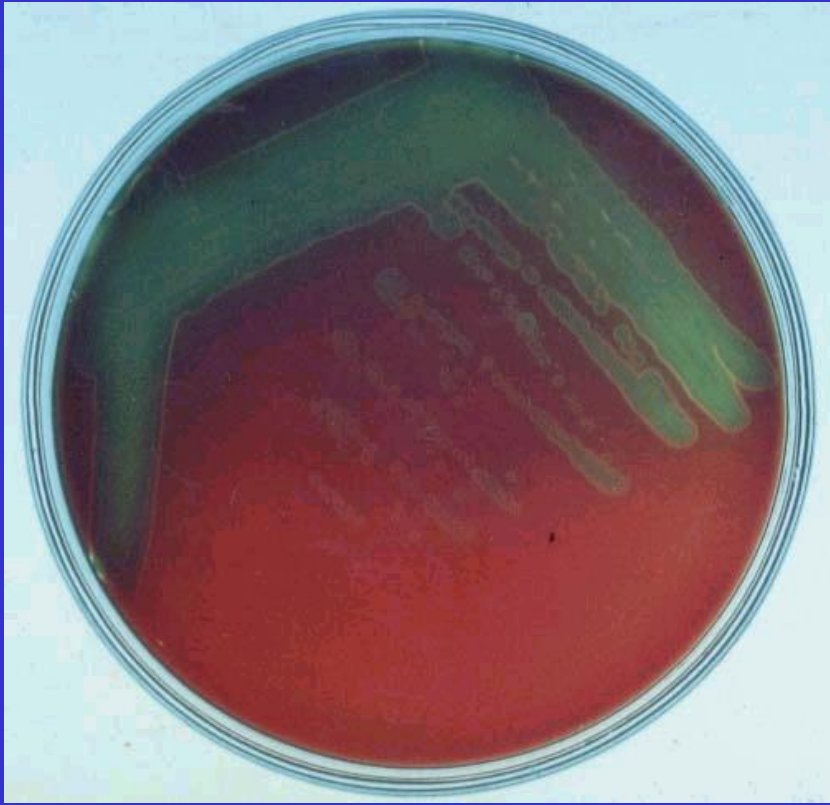


5. *Streptococcus pyogenes* on blood agar



Pinpoint colonies,
strong β -hemolysis!

6. *Str. mitis* on blood and chocolate plates



7. *Streptococcus pneumoniae* on blood agar plate (optochin S)



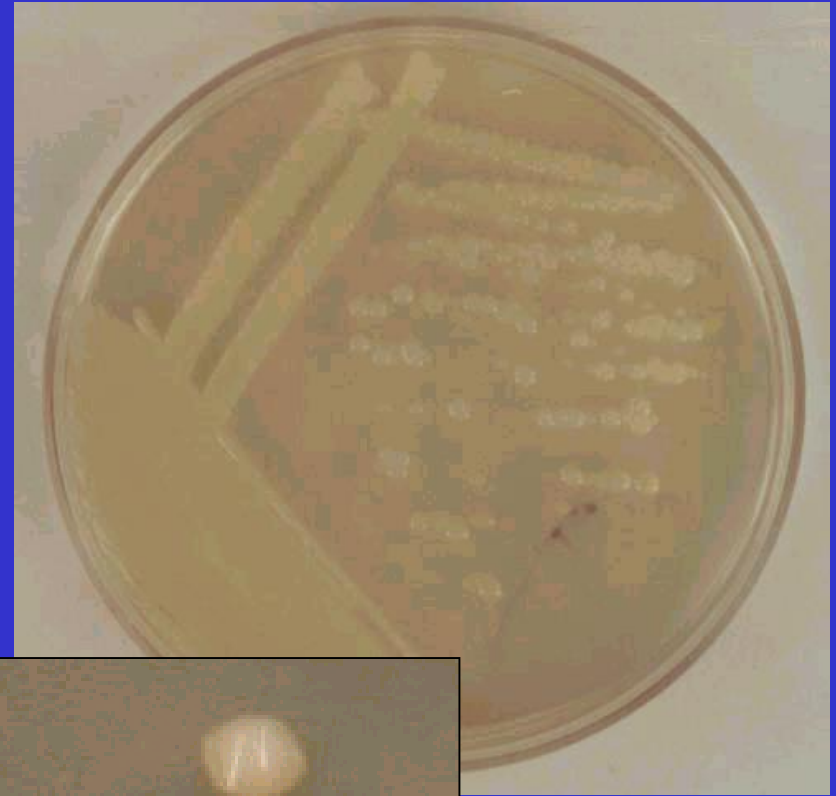
8. *Haemophilus influenzae* on chocolate agar



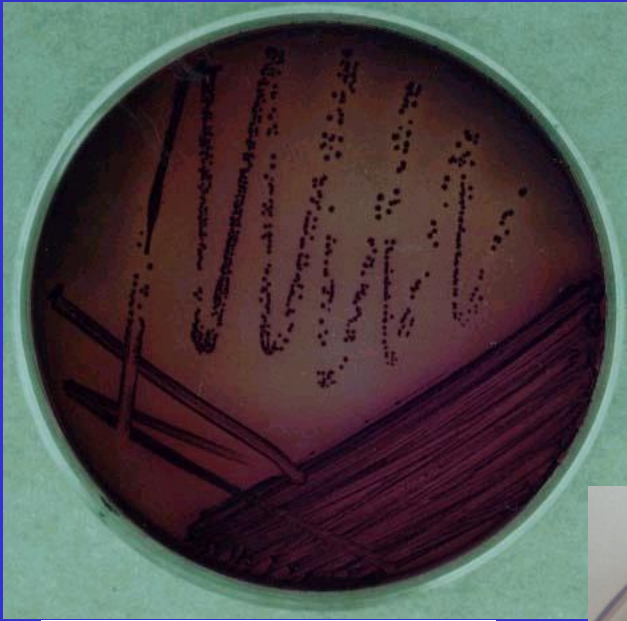
„Morning dew” colonies,
running nose smell!



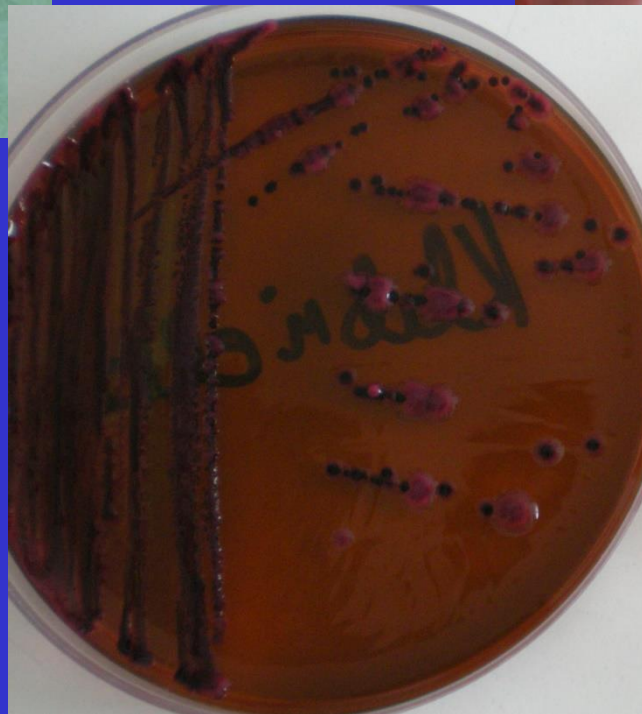
9/12. *E. coli* and *Klebsiella* on agar plate



9/12. *E. coli* and *Klebsiella* on EMB plate



M022
Escherichia coli (25922)



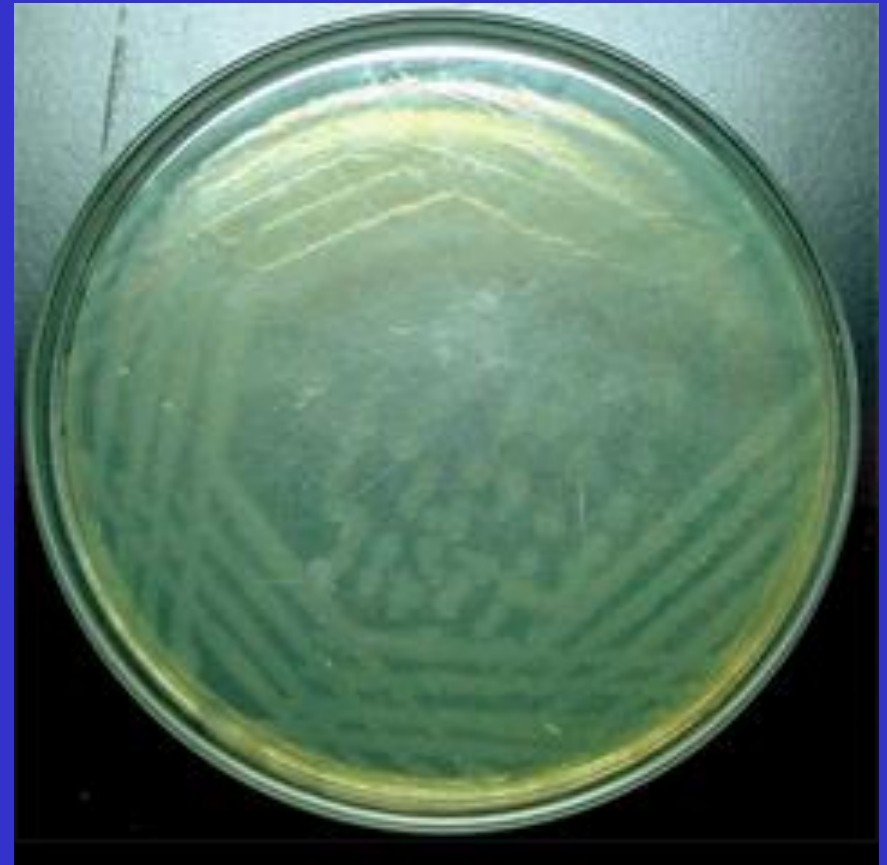
10. *Proteus* on agar and blood agar (swarming!)



11. *Pseudomonas* on agar plate



Pseudomonas aeruginosa

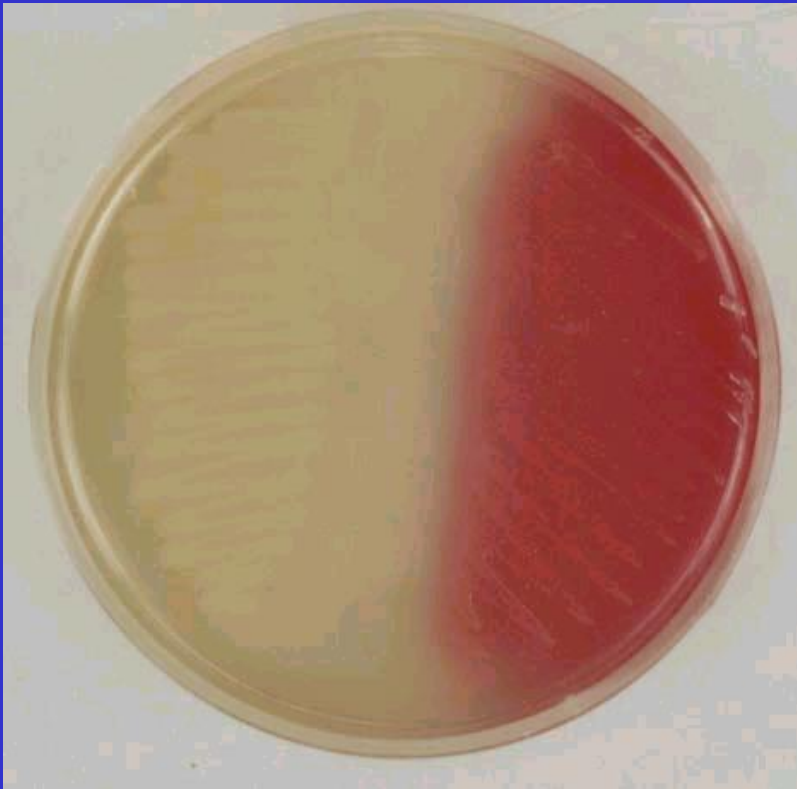


- Green pigment!!!
- Pleasant odour
- Mucoid colonies

13. *Salmonella* on bismuth sulphite medium



14. *E. coli* and *Salmonella* on brilliant green medium



15. *E. coli* and *Shigella* on DC medium



16. Faeces of patient with dysentery on DC medium (*E. coli* + *Shigella*)



18. Urease test

- $\text{NH}_2\text{-CO-NH}_2$ (ureum) $\rightarrow$ $\text{CO}_2 + \text{NH}_3$ (ammonia)
- Christensen medium:
indicator (phenol red)
 - urease +: purple
 - urease -: citric yellow

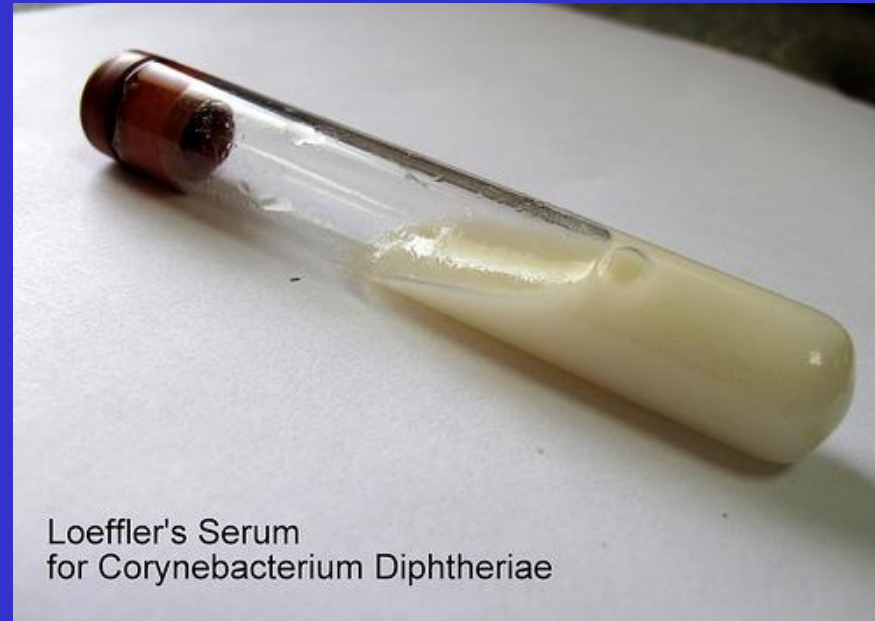


Urease +: *Proteus, Klebsiella, Helicobacter*

19. *Corynebacterium* on Clauberg and Löffler medium



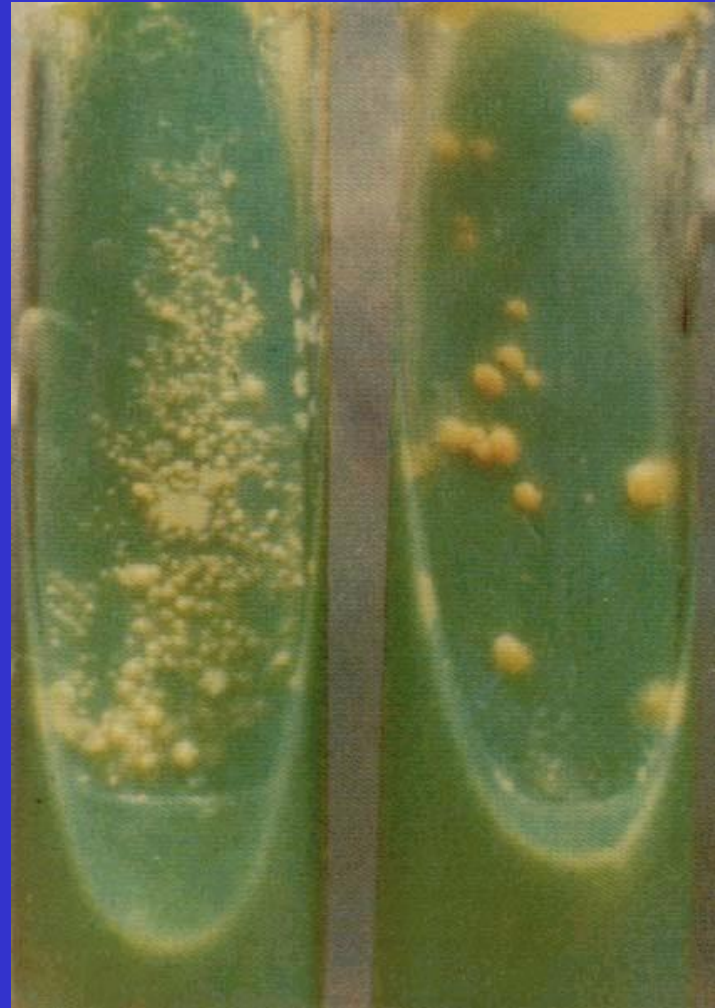
Clauberg



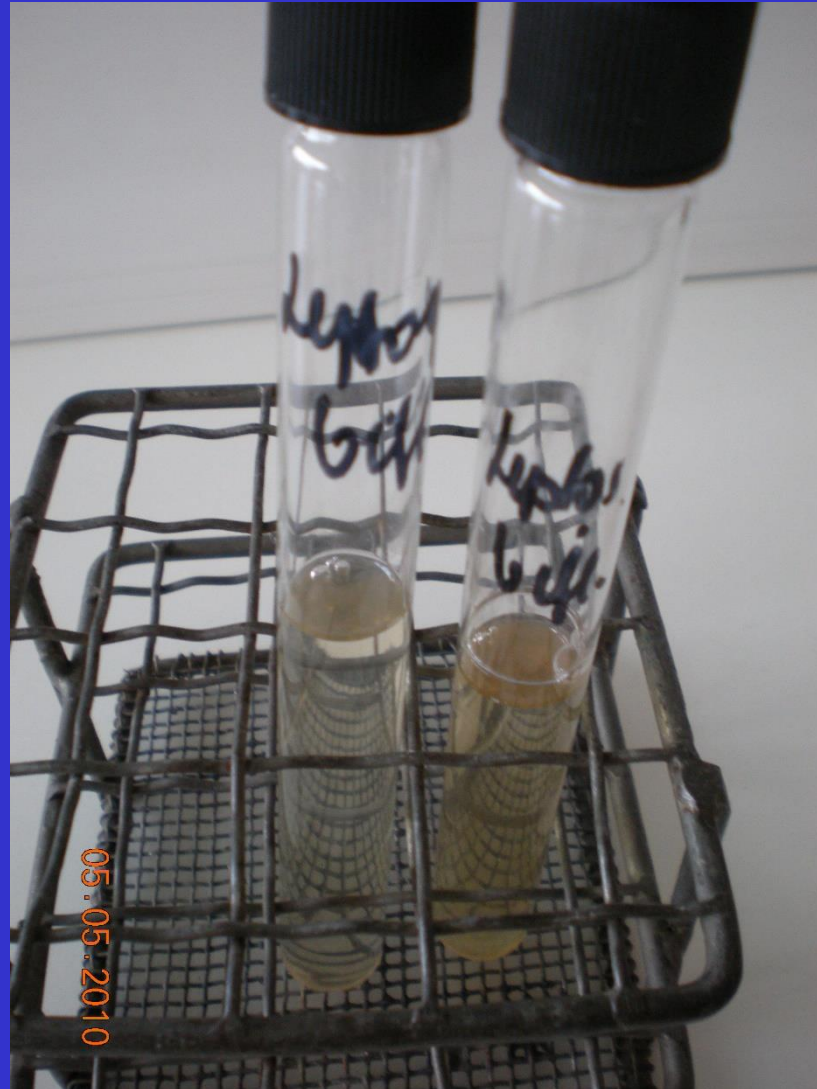
Loeffler's Serum
for *Corynebacterium Diphtheriae*

Löffler

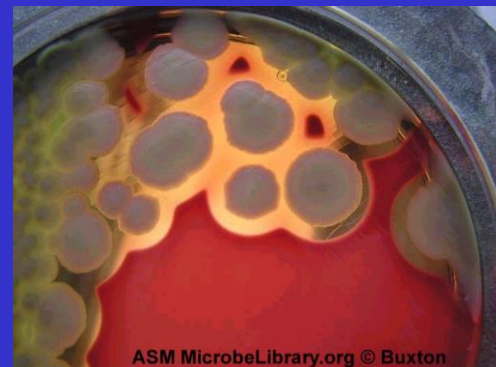
20. *Mycobacterium tuberculosis* on Löwenstein-Jensen medium



21. *Leptospira* in Korthof medium



22. *Bacillus cereus* on agar and blood agar plate



23-24. *Clostridium tetani* and *C. perfringens* in Holman and thioglycolate media



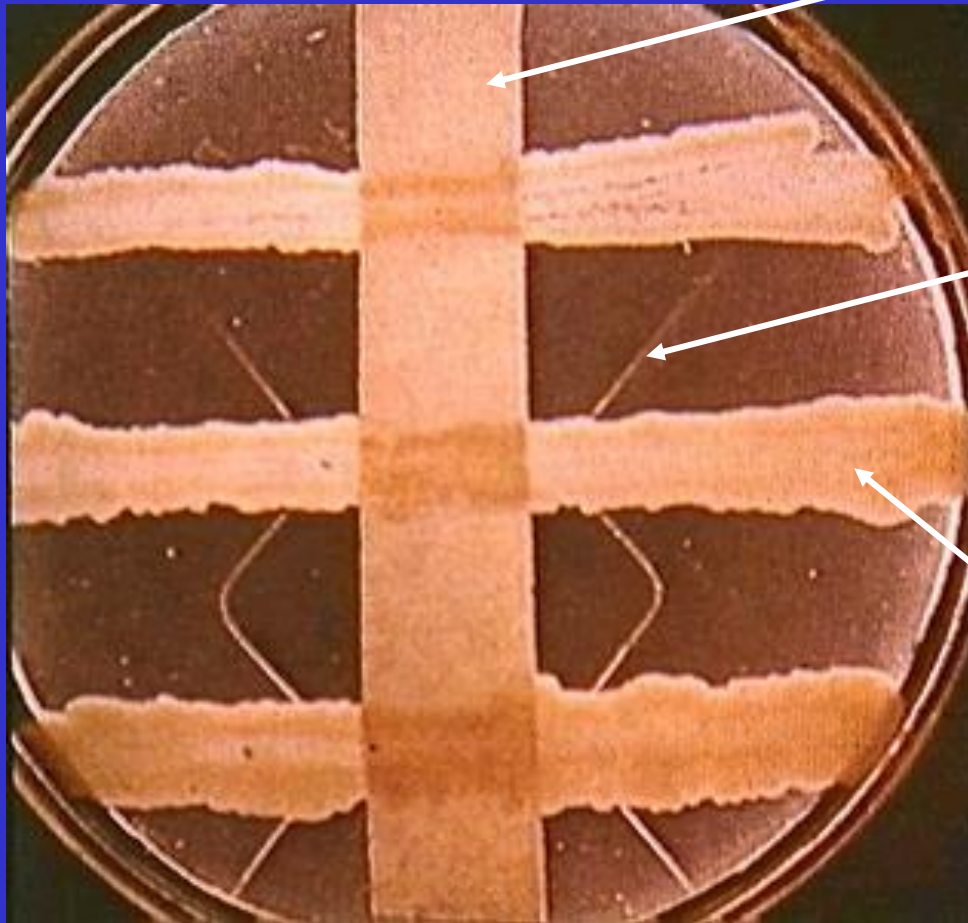
GasPak



III. Reactions and methods

1. Elek test

Aim: test for toxin production



Filter paper
with antitoxin

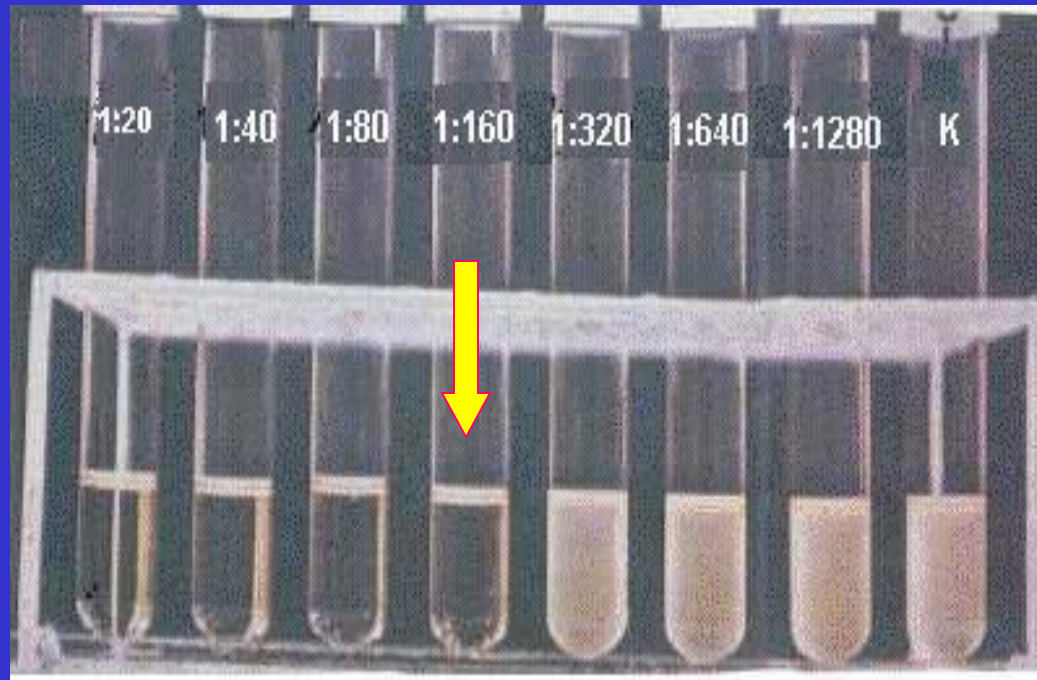
Precipitation
arch

Corynebacterium

2. Widal-type reaction

- Gruber-Widal: abdominal typhus (*Salmonella*)
- Weil-Felix: epidemic typhus (*Rickettsia*)
- Wright: *Brucella* / *Francisella*

With
Proteus
OX19
antigen!

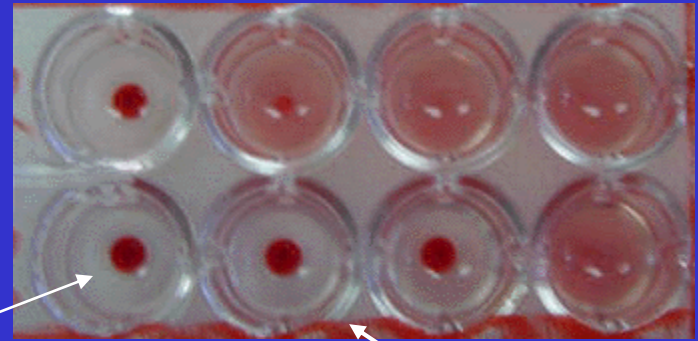


titre: the smallest amount of antibody, which can cause positive reaction!!!

3. Wasserman test for syphilis (CF)

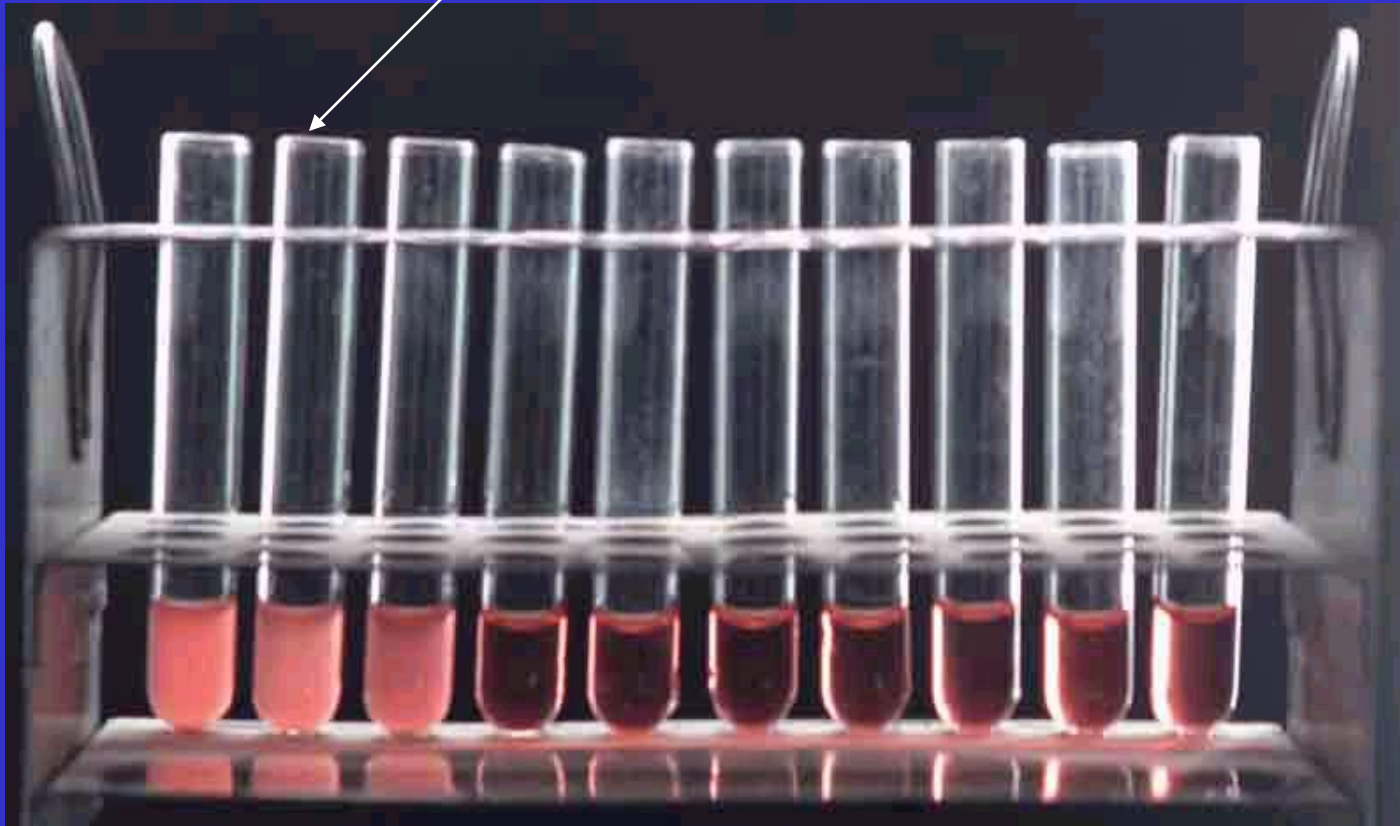
1

2

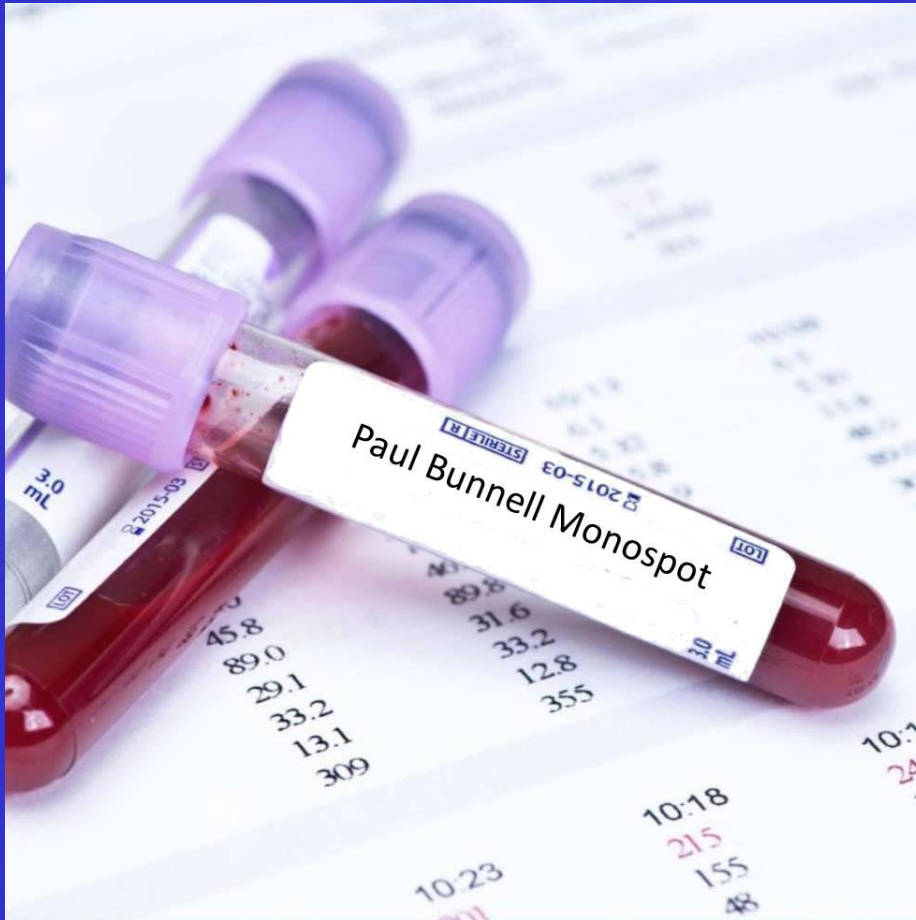


Positivity: no hemolysis

2: 4-fold increase in titre compared to **1** - indicates infection



Paul-Bunnell test

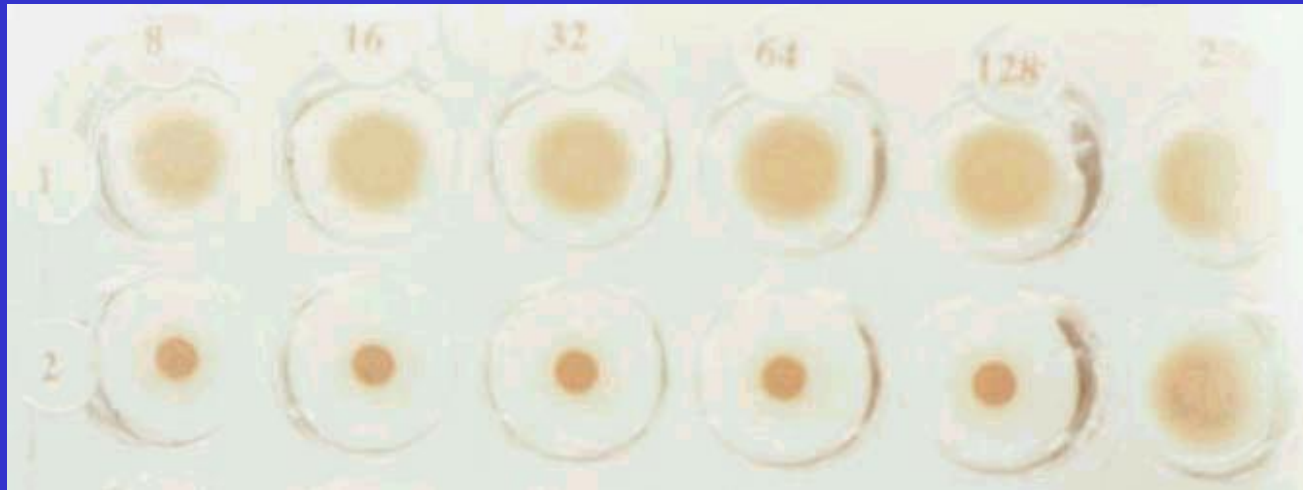


- Mononucleosis infectiosa
- EBV: P-B positive
- CMV: P-B negative
- Patient's antibodies agglutinate sheep rbc

3. Hemagglutination inhibition (HI) (Rubella)

NEG K	POZ K	A savópár 1. minta	A savópár 2. minta	B savópár 1. minta	B savópár 2. minta	B savó 2. minta <u>össz-</u> ellenanyag	B savó 2. minta IgM eltávolítás
1:320							
1:160							
1:80							
1:40							
1:20							
1:10							

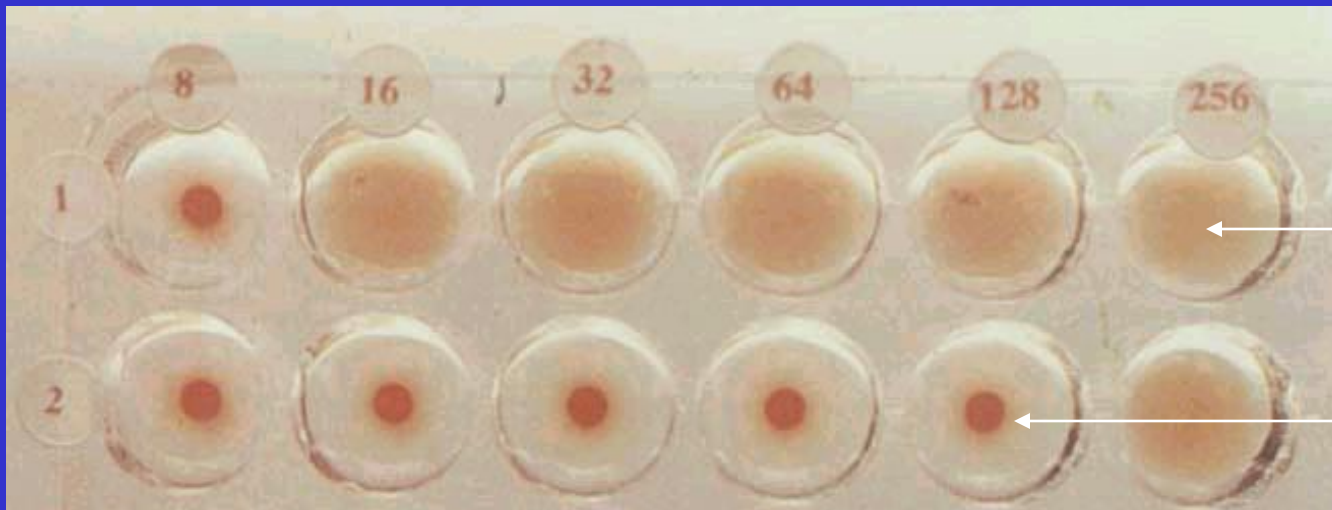
4. Hemagglutination (HA) and -inhibition (HAI)



hemagglutination

hemagglutination
inhibition

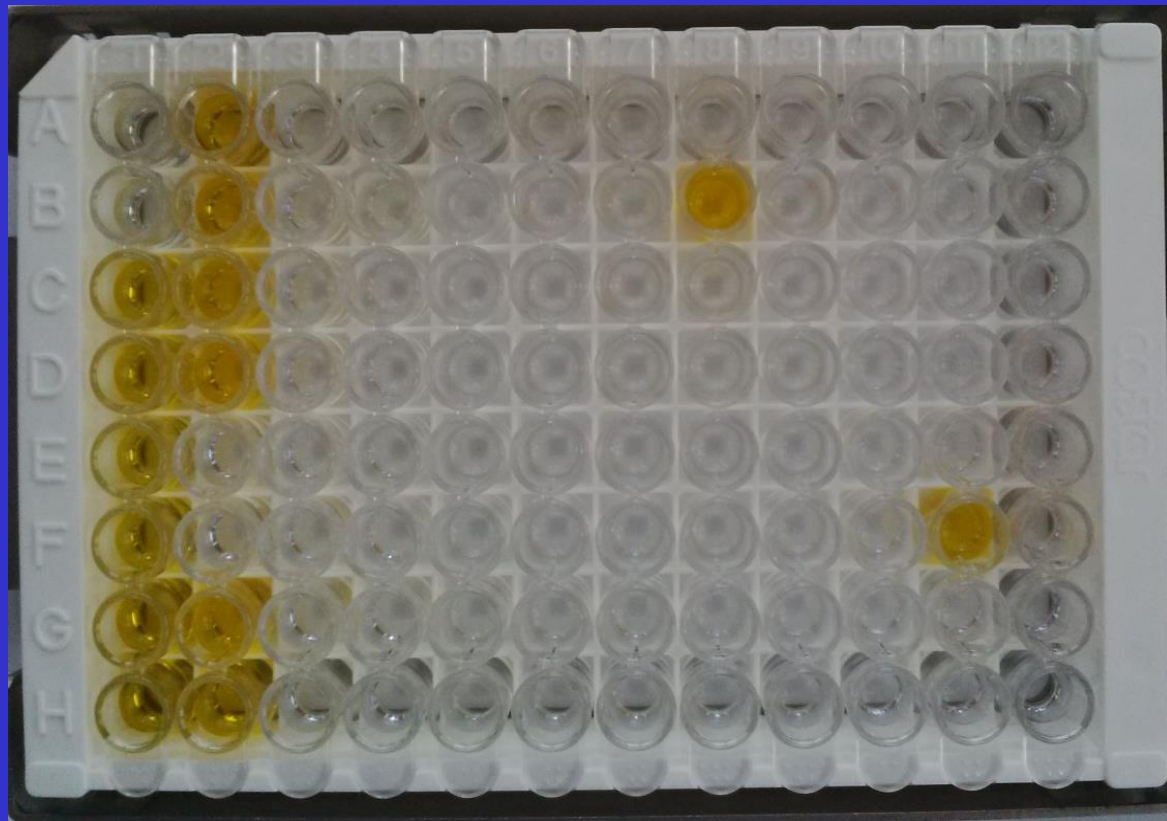
CF test



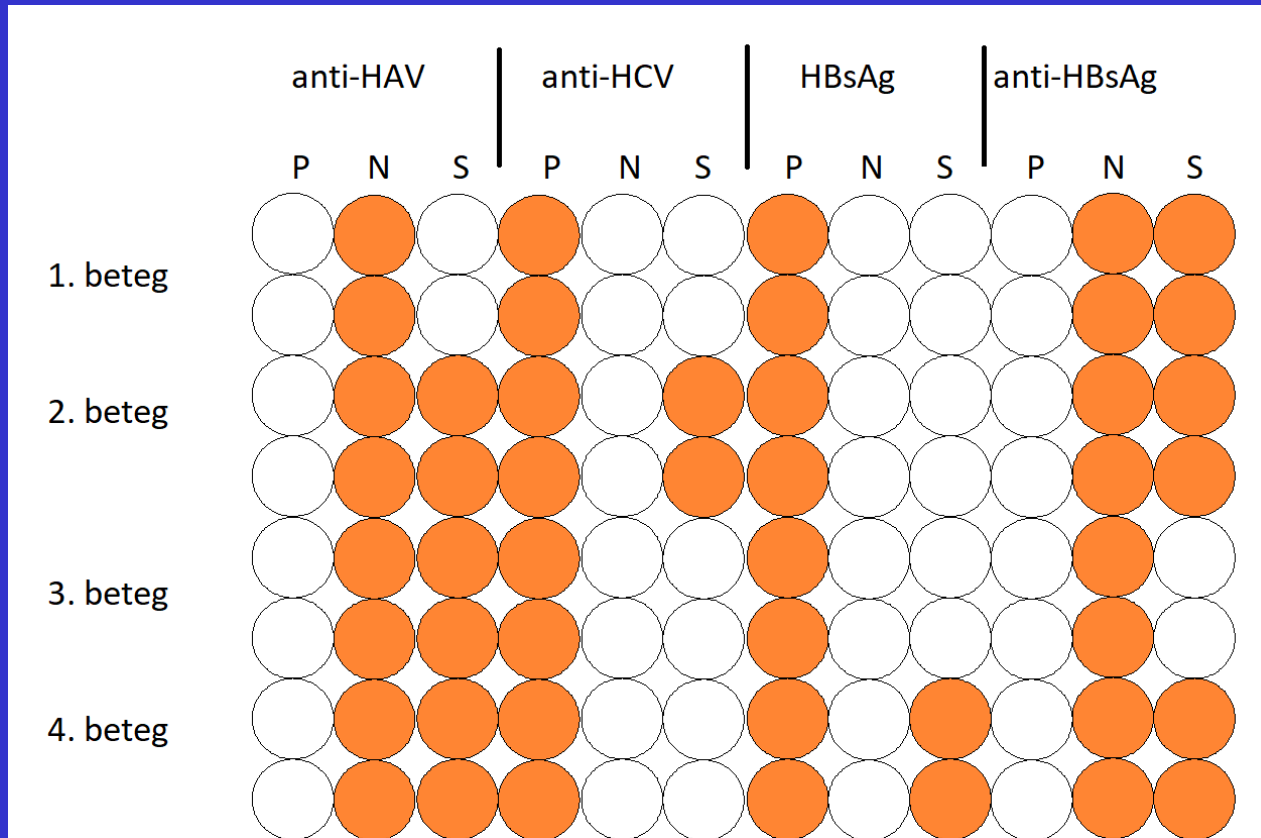
hemolysis (-)

no hemolysis (+)

5. ELISA tests: HIV-1 and HIV-2



5. ELISA tests: Hepatitis



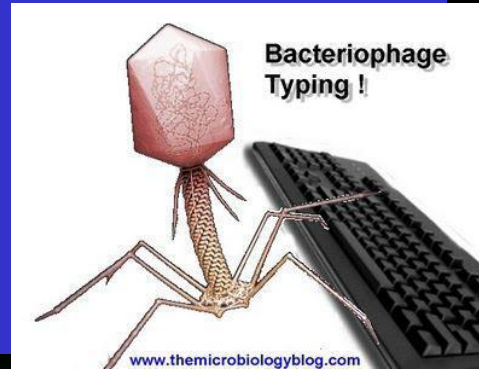
Anti-HAV
(competitive ELISA)

Anti-HCV (indirect
ELISA)

HBsAg (sandwich
ELISA)

Anti-HBsAg
(competitive ELISA)

8. Phage typing



Phage type
52-52a-80

	90		92	
29	52	52a	79	80
3a	3c	55	71	187
6	42e	47	53	54
75	77	83a	84	85
95	96	88	81	42d



10. Steps of Gram staining

1/a. Crystal violet 2'



1/b. Lugol solution 1'



2. Differentiation with 96 % ethanol

3. Washing with water

4. Safranin 1'



5. Washing

6. Drying

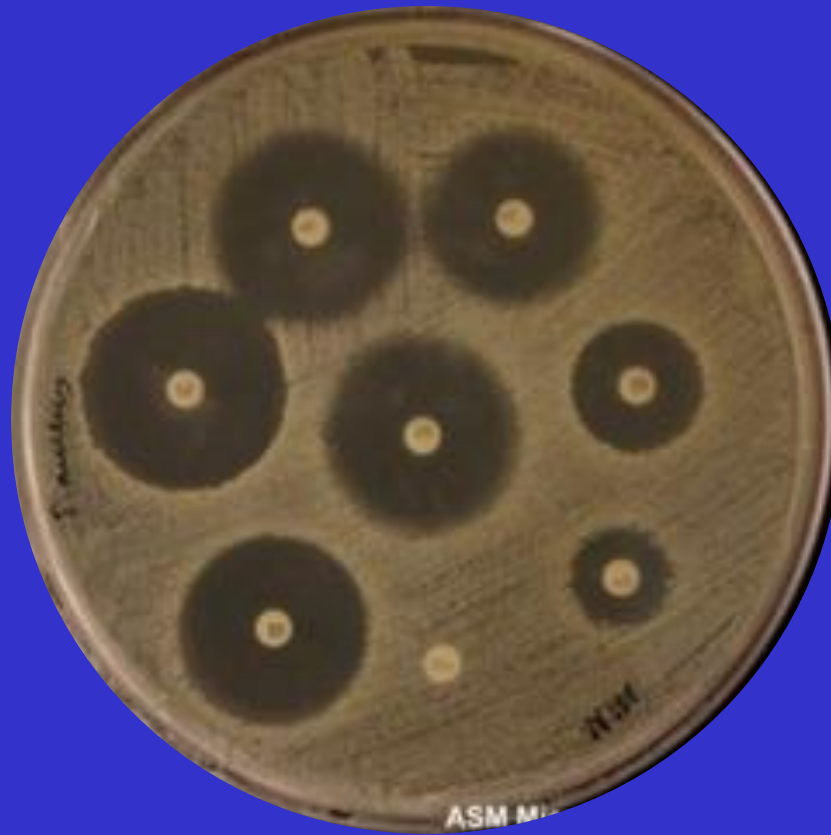
7. Microscopic examination (immersion)

11. Steps of Ziehl-Neelsen staining

“acid fast staining”

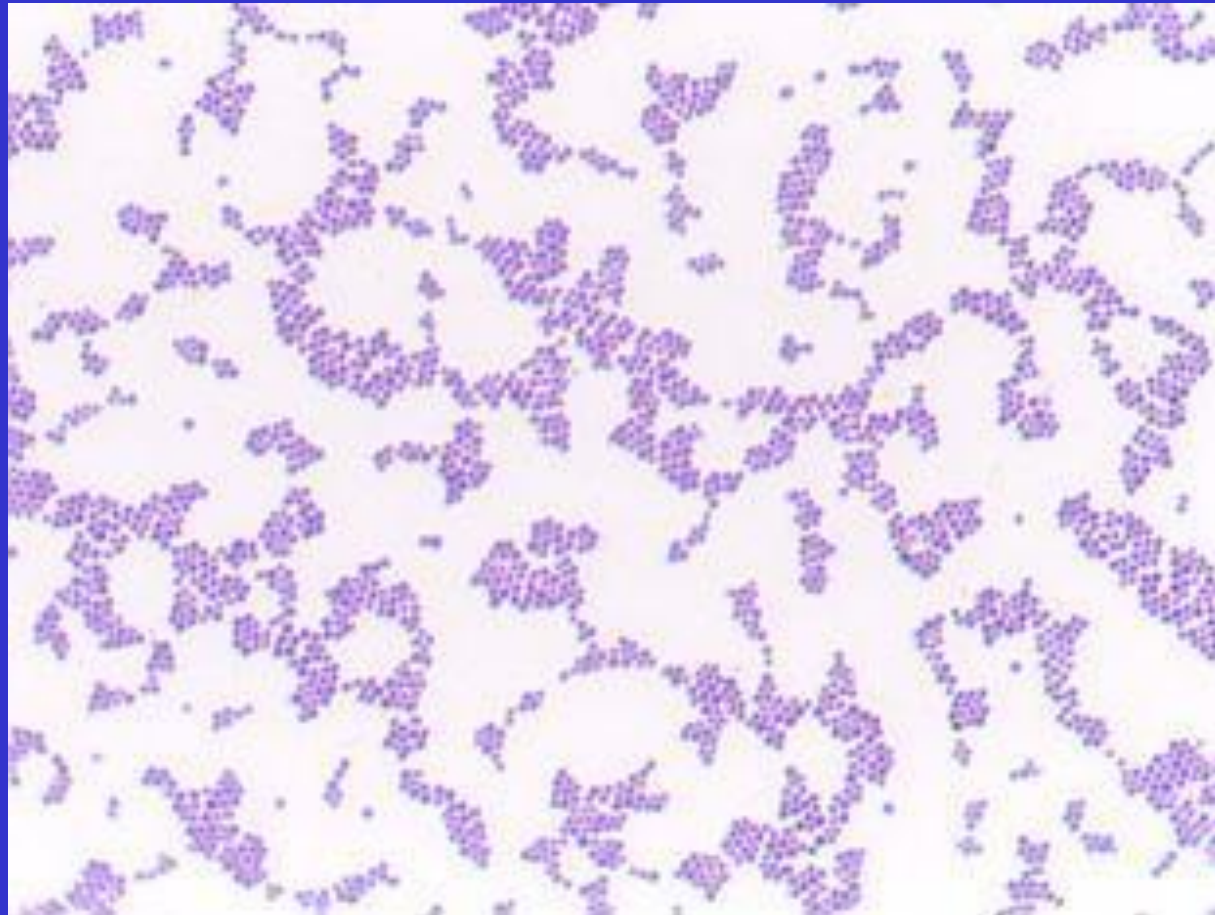
- Carbol-fuchsin (on filter paper!)
 - heating 3x (until steaming)
- Differentiation: acidic alcohol
 - 3% HCl, 96% ethanol
 - “acid fast”
- thorough water washing
- counter staining 1-2’
 - methylene blue or malachite green
- careful rinsing, drying with filter paper

Antibiotic susceptibility test with disc diffusion (“antibiogram”)

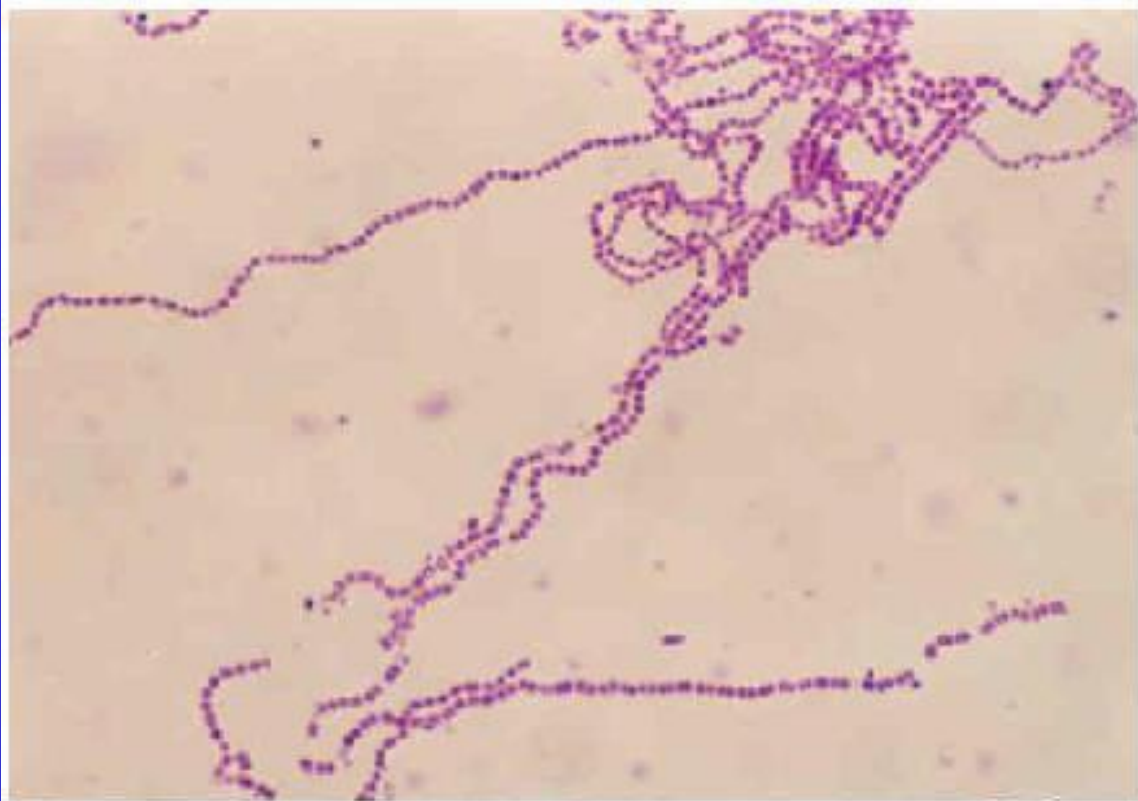


IV. Microscopic preparations

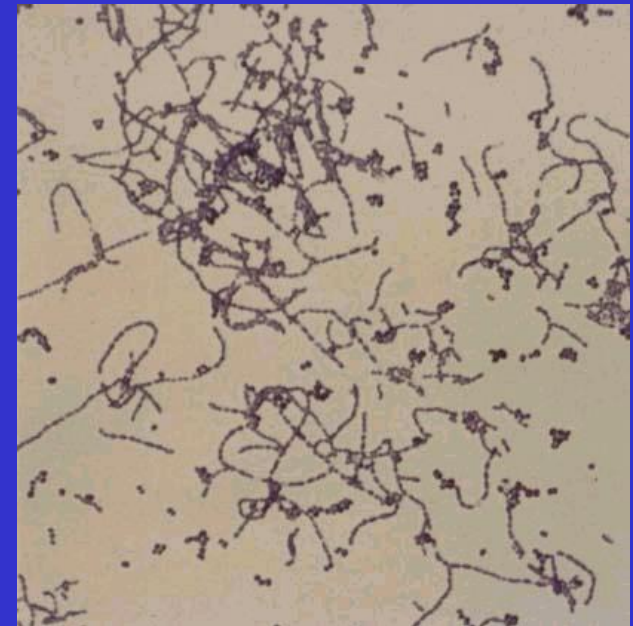
1.a *Staphylococcus* (Gram +)



1.b *Streptococcus pyogenes* (Gram +)



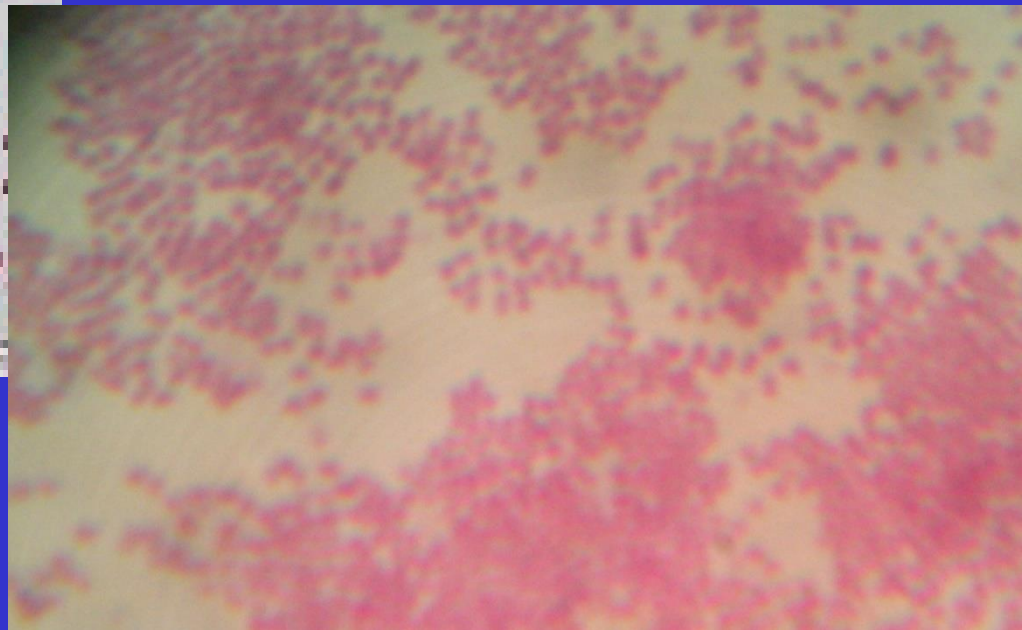
Gram pozitiv coccus (*Streptococcus pyogenes*)



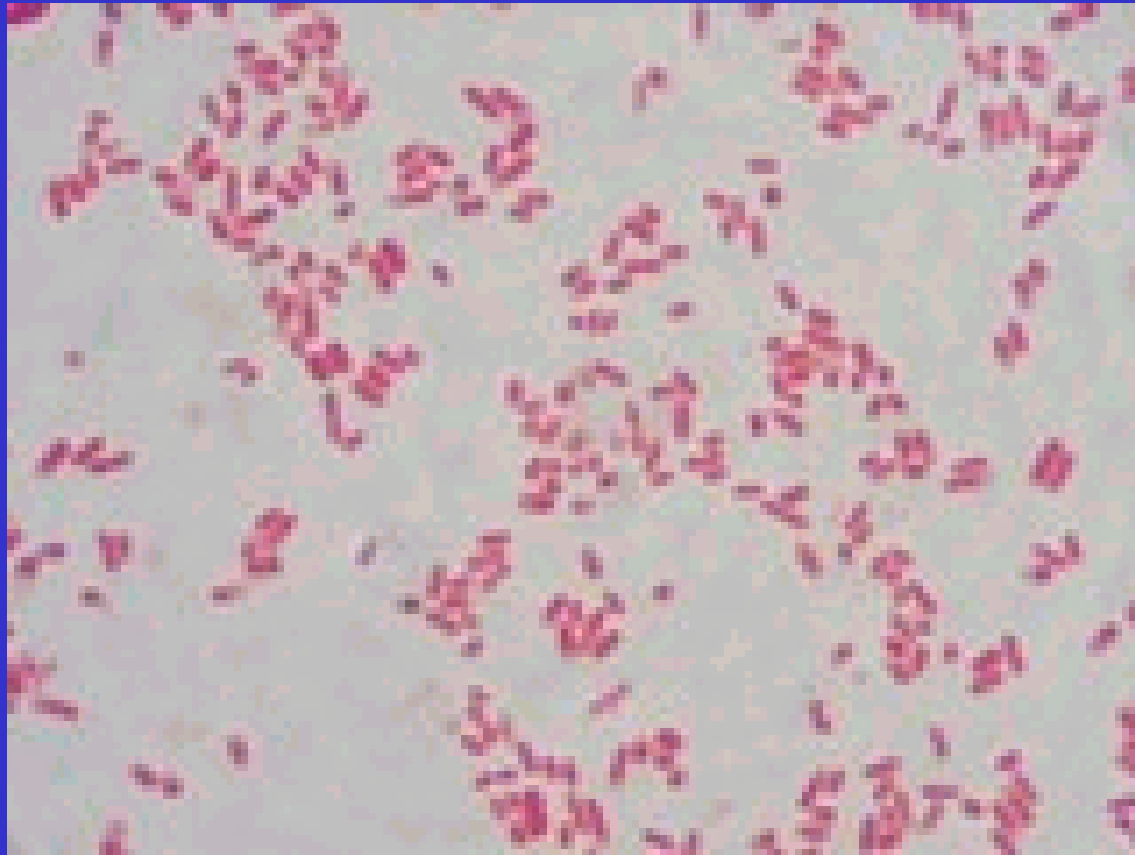
1.c *Streptococcus pneumoniae* (G +)



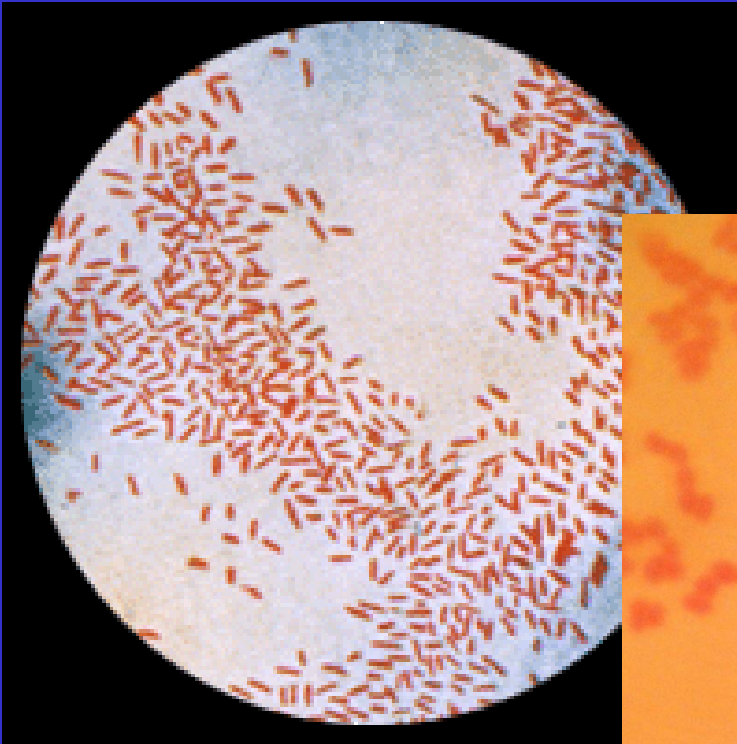
1.d *Neisseria* (Gram -)



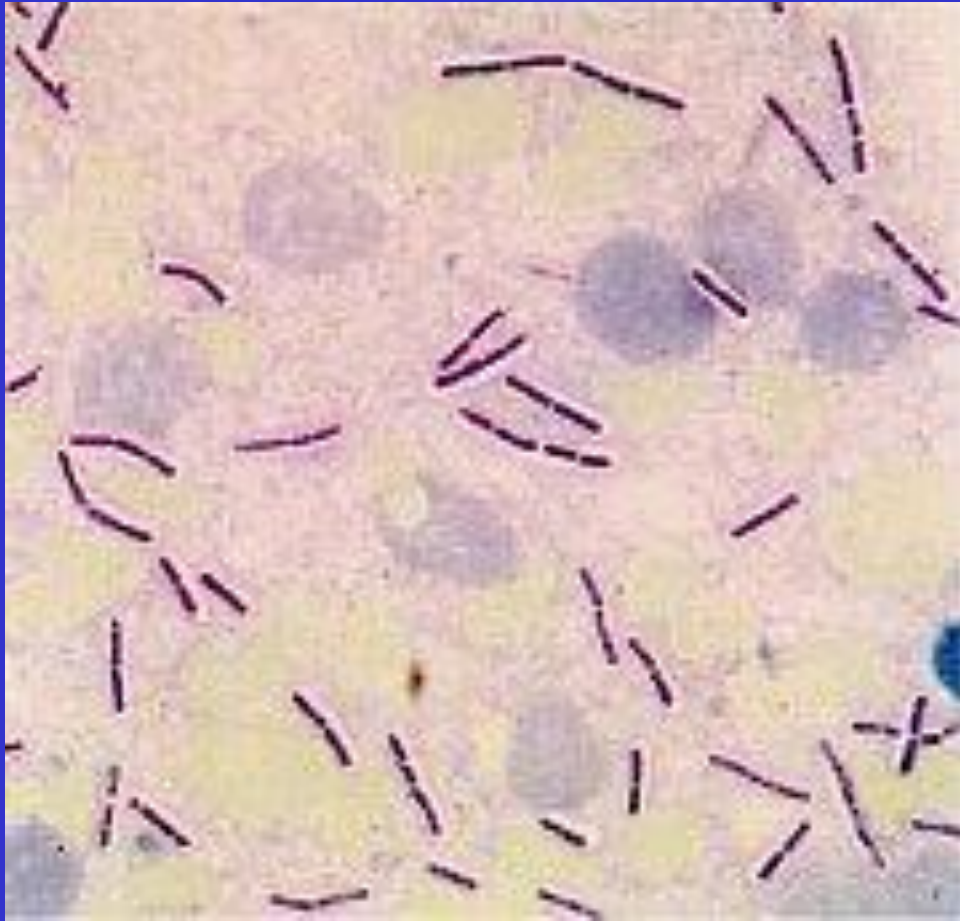
1.e *E. coli* (Gram -)



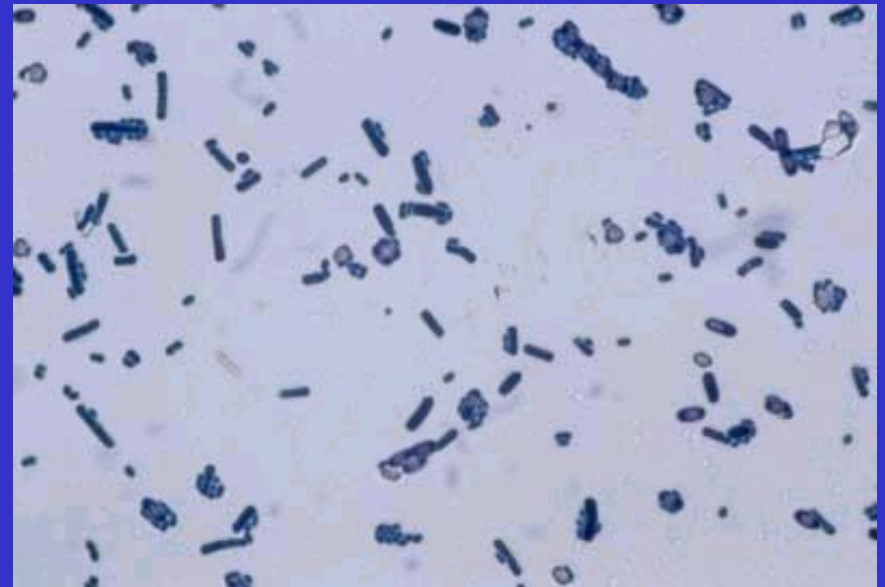
1.f *Haemophilus influenzae* (Gram -)



1.g *Bacillus* (Gram + spore forming)



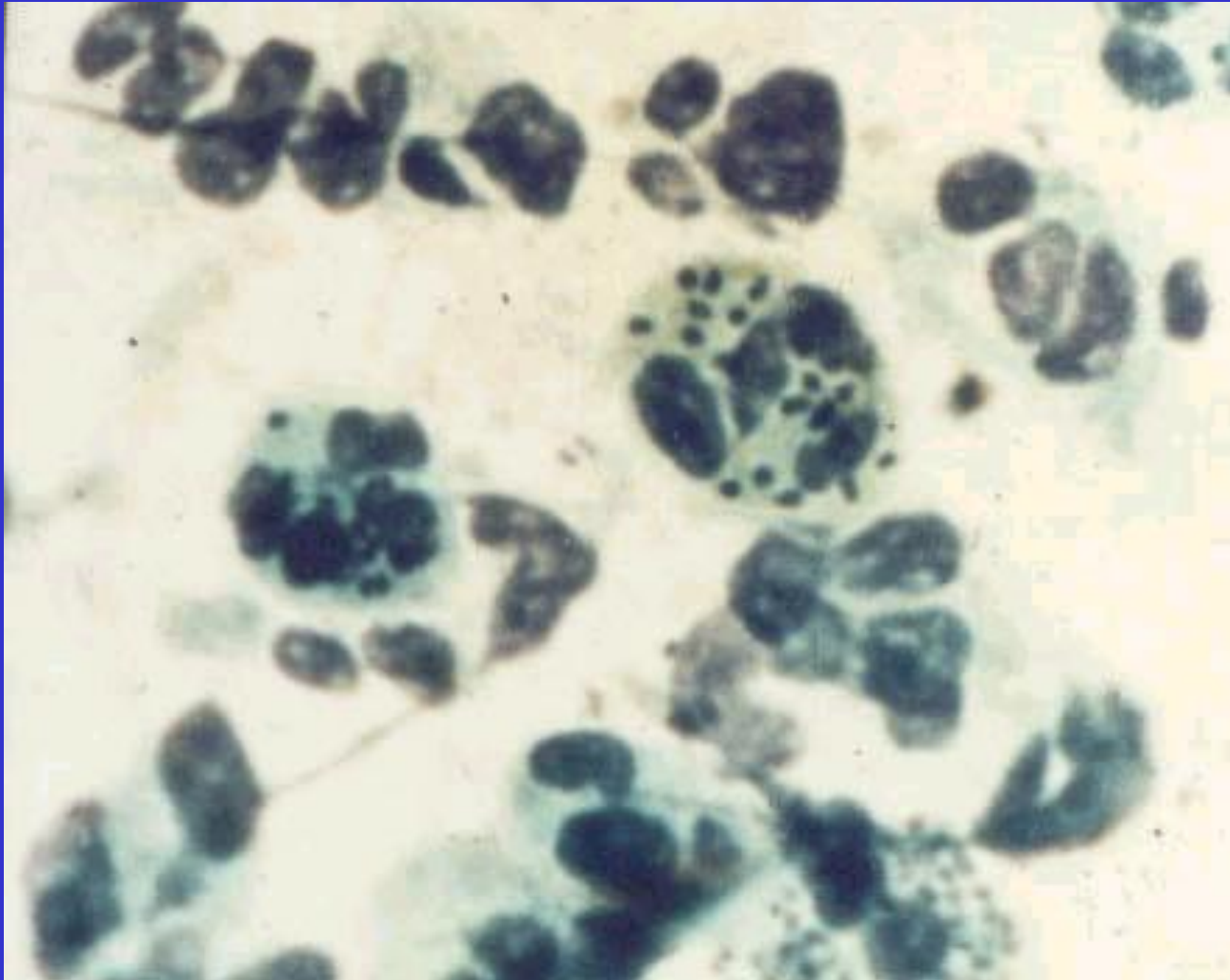
1.h *Clostridium tetani* and *perfringens* (Gram + spore forming)



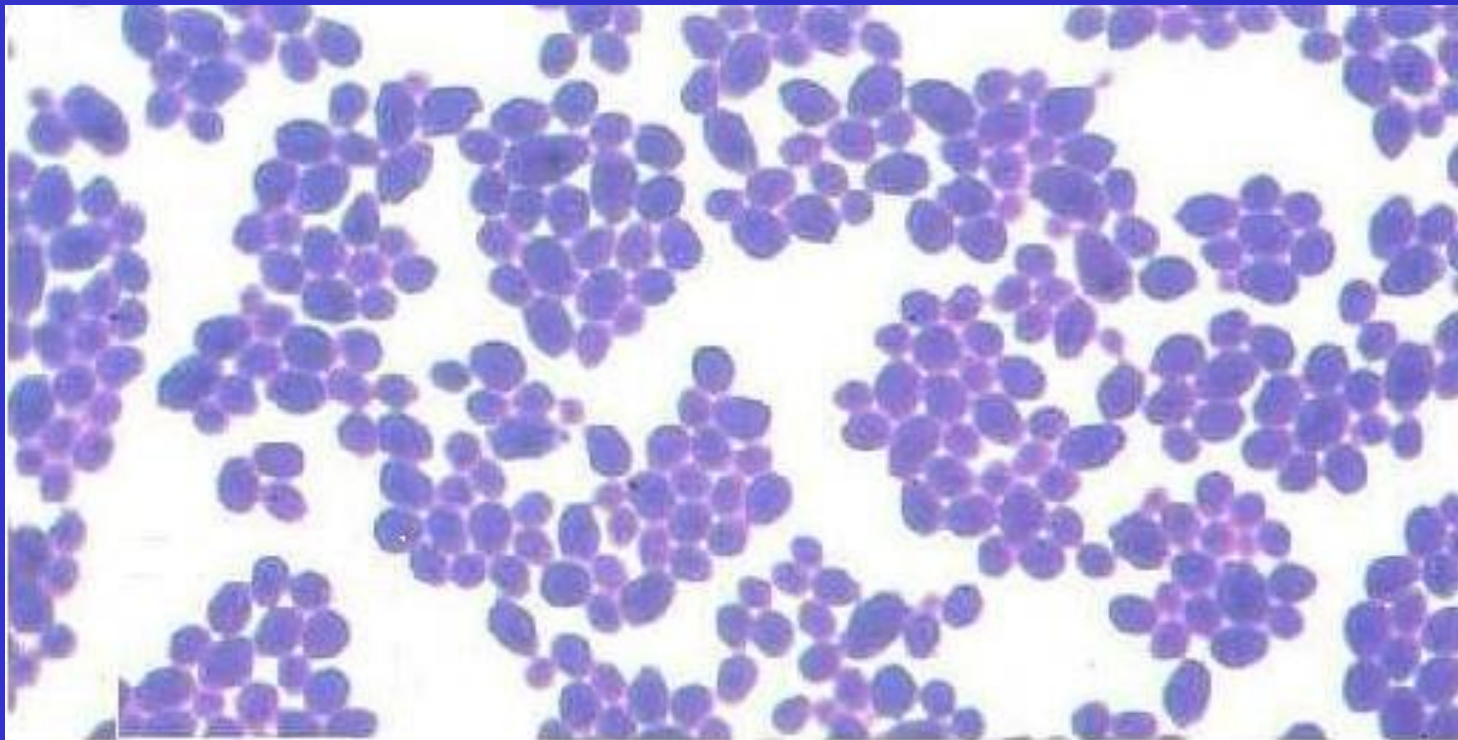
1.i *Corynebacterium* - Neisser staining



2.a *Neisseria gonorrhoeae* in pus (methylene blue staining)



2.b *Candida albicans* from culture

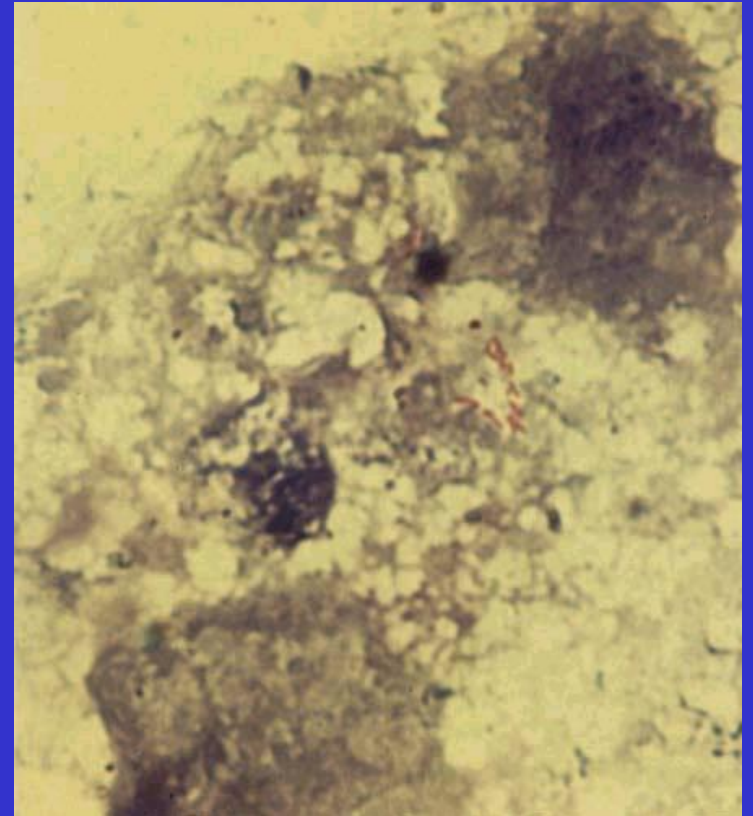
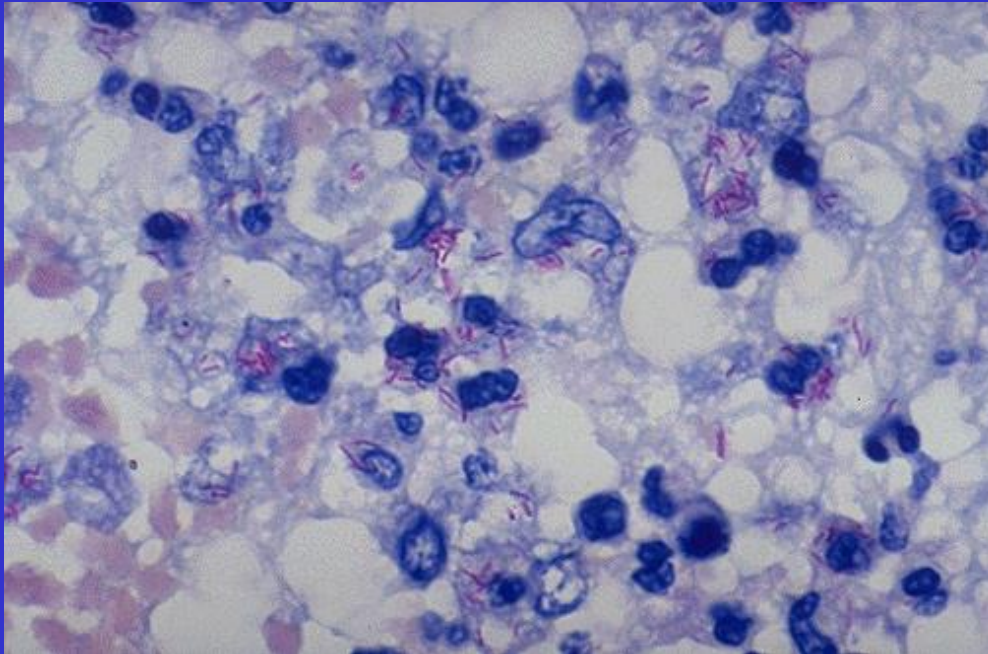


Sarjadzó gomba (*Candida albicans*) egyszerű festéssel (metilénkék) készített kenete

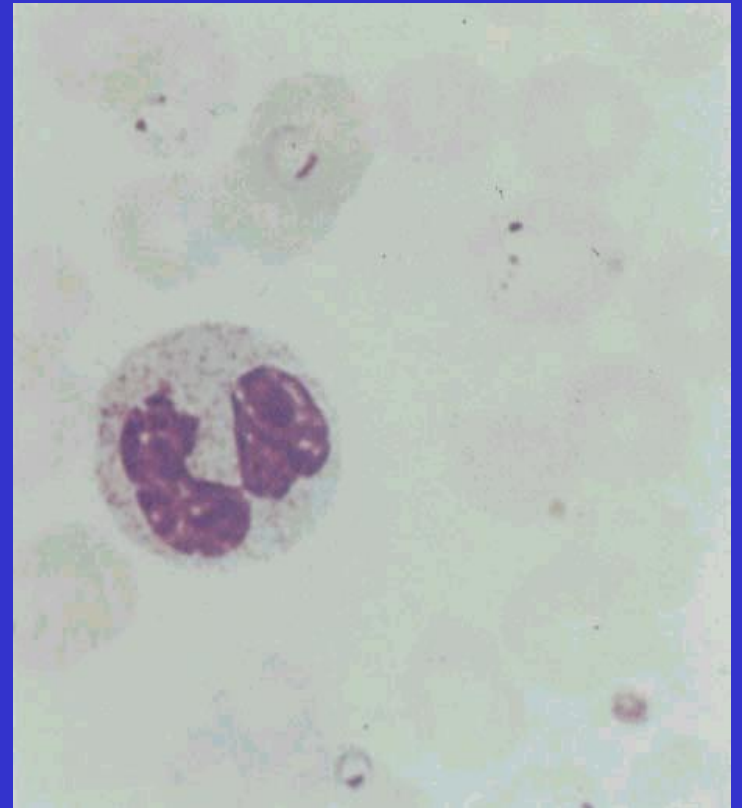
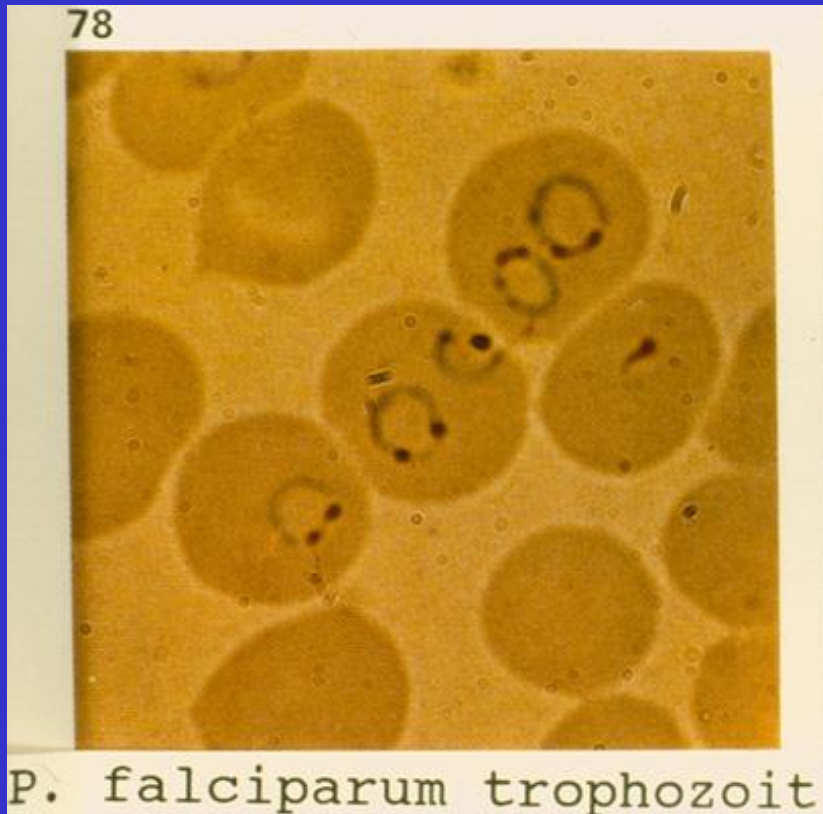
3. Silver impregnation: *Leptospira*



4. Ziehl-Neelsen staining: *Mycobacterium tuberculosis*



5.a *Plasmodium falciparum* in thin blood film (Giemsa staining)



5.b *Trypanosoma brucei* in thin blood film (Giemsa staining)



6. Anorectal cellulose-tape preparation (*Enterobius*)



VI. Tests performed by the students

Catalase test

Catalase +: *Staphylococci*

Catalase -: *Streptococci*

- **performance**: adding a few drops of H_2O_2 to the culture, strong bubbling if positive
- $\text{H}_2\text{O}_2 \rightarrow \text{H}_2\text{O} + \text{O}_2$



Clumping test - Coagulase test

Coagulase +: *Staphylococcus aureus*

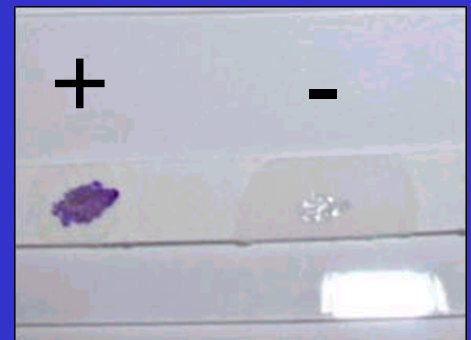
Coagulase -: all other *Staphylococci* (“CNS”)

- “Staphaurex”: latex agglutination
- **performance**: mixing bacteria with fibrinogen-bound latex particles on microscope slide → **clumping**

Oxidase test

Oxidase +: *Pseudomonas*, *Neisseria*,
Vibrio, *Campylobacter*

- **purpose**: detection of cytochrome-oxidase enzyme
- diagnostic use: differentiation of obligate aerobes
- **performance**:
 - filter paper on microscope slide
 - add oxidase reagent
(= tetraphenyl-pyrazine derivative)
 - add bacteria
 - **positivity**: pink / purple colour



Indol test

- triptophane → indol
(tryptophanase enzyme)
- amylalcoholic (=organic)
reagent → indol is dissolved
here after mixing → intensive
red ring on top (alcohol is
lighter than water!)
- can be performed in ureum-
indol tube

Indol +: *E. coli*



Indoltermelés kimutatása:

a peptonvizés baktériumtenyészetet amilalkoholban oldott sósavas
paradimetil-amino-benzaldehiddel összerázva pozitív esetben az
amilalkoholos fázis élénk vörös színű lesz

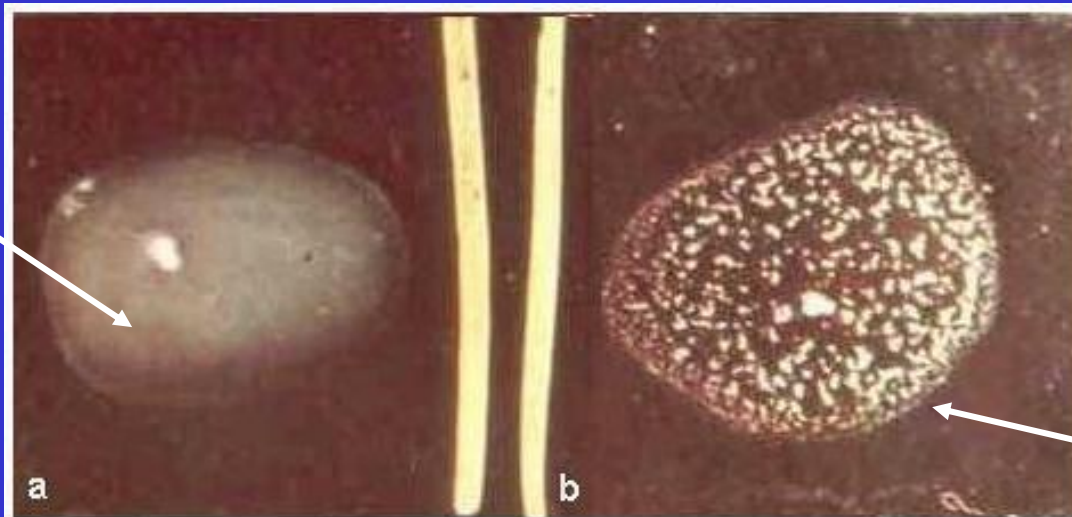
1. kontroll
2. negatív (*Klebsiella*)
3. pozitív (*E. coli*)

6. Slide agglutination with *E. coli*

NEGATIVE

POSITIVE

Bacterium
suspension
(control)



Tárgylemez agglutináció (szerotipizálás, kvalitatív reakció, a: negatív, b: pozitív, rögzös kicsapódás)

White
particles,
clear
solution