Semmelweis University Department of Laboratory Medicine

Founded: 2010

- Major profile: diagnostics
 - 4.5 Million tests pro year
 - 450,000 patients
- Up to 65 per cent of lab investigations performed at Semmelweis University + national centre for special tests
- Education: one semester, 14 lectures started in 2010

Informations on the Institute

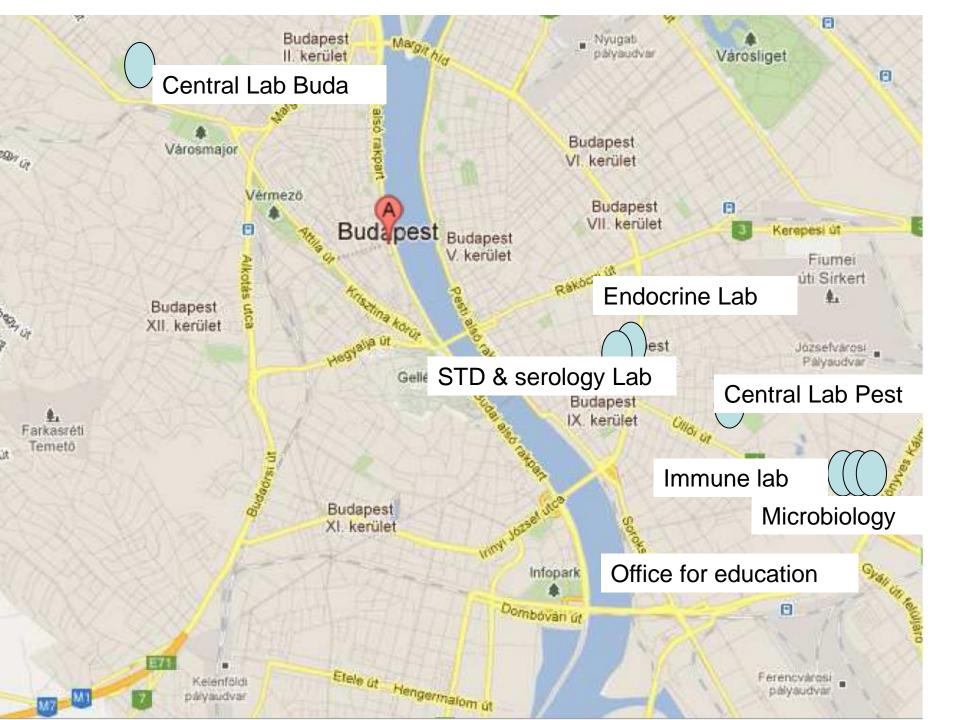
Office: Nagyvárad square, Floor 14.

Head: prof. Barna Vasarhelyi, DSc

Contact:

vasarhelyi.barna@med.semmelweis-univ.hu

For further info: semmelweis.hu\laboratorium



Some words about this discipline

- Lectures
- Supporting materials

Clinical disciplines **LABORATORY MEDICINE** Microbiology **Pathophysiology**

Exam

- Test exam (single choice)
- See detail on the homepage

Factors having an effect on lab test results



Result interpretation

Test request





Sampling



Black box: the lab



Medical care

The cumulative number of fatalities caused by

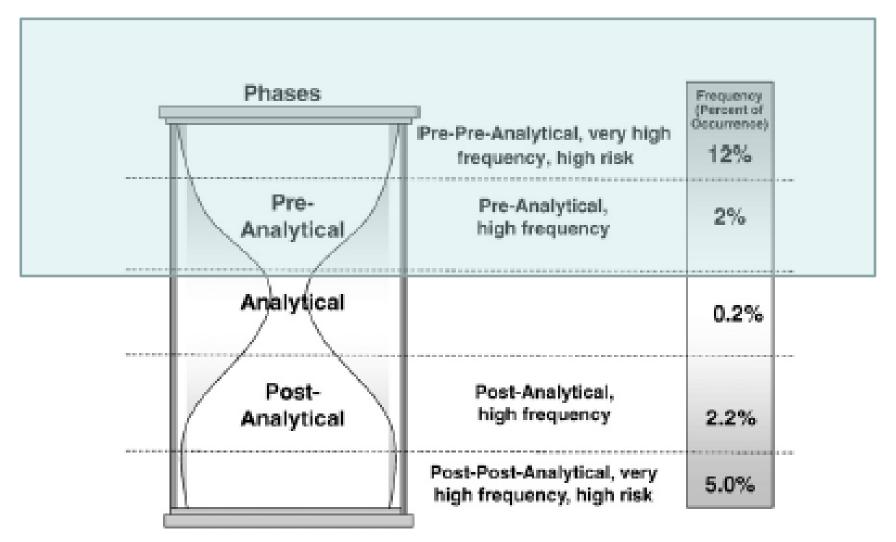
- Accidents
- Breast cancer
- HIV infection

is LOWER, than those caused by maltreatment (US figure).

Estimated number: 50 – 100 thousand per year Majority of cases are due to diagnostic error. The 70% of diagnostic tests are performed in the lab. Prevalence of incorrect results:

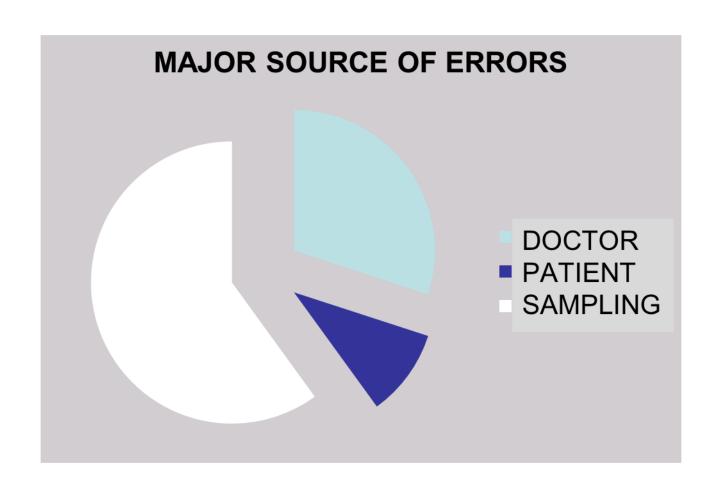
1/ 164 - 1/330 measurements

OCCURRENCE OF FAULTS WITH LAB TESTS



Phase	Percentage of all missed diagnosis
Pre-Analytic	
-Failure to order appropriate diagnostic or laboratory tests	55
-Adequate diagnostic or laboratory tests ordered but not	
performed	9
Analytic	
-Diagnostic or laboratory test performed incorrectly	8
Post-Analytic	
-Incorrect interpretation of diagnostic or laboratory tests	37
-Responsible provider did not receive diagnostic or	12
laboratory test results.	

THIS LECTURE IS ABOUT PREANALYTICAL PHASE (AND ERRORS DONE)



Major factors responsible for preanalytical variations

DOCTOR:

1. Indication of lab test

PATIENT:

- 1. Biological variability
- 2. Environmental factors

SAMPLING:

- 1. Patient ID
- 2. Patient preparation
- 3. Sampling and transport

Should know:

What (what kind of test)

What for (the goal)

When (or how frequent)

Source of specimen (blood, urine)

Destination (lab the sample to be sent)

Costs (...)

Why does somebody ask lab test?

- Esetablish the diagnosis
- Monitoring the therapy / condition
- Screening (exclusion of a disease)

The parameter to be tested depends on the goal [see: lecture on postanalytics]

Factors responsible for preanalytical errors - DOCTOR When is a lab test justified?

If it adds significant info supporting the clinical decision making

Component	Note	Contraindication (example)
significant	ie: information essential for the decision making	Requesting vitamin D-levels in infection
adds	le: data available just by this way or by another more expensive /invasive test or	Cholesterol levels in acute infarction
Decision making	Ie: it is time to decide anything]	Tumor marker levels in terminal phase of malignancy

What does one expect from a lab test?

- To be as specific and as sensitive as it is possible. Discriminate the disease from the healthy condition or recognize condition requiring intervention
- An appropriate marker should be selected.

Marker: what does one expect from a biomarker?

- Easily accessible (e.g. present in blood)
- Easily measured (does not require specifif machine)
- Stable and easy to repeat
- Influencing factors are known
- Reference range is known
- Cheap

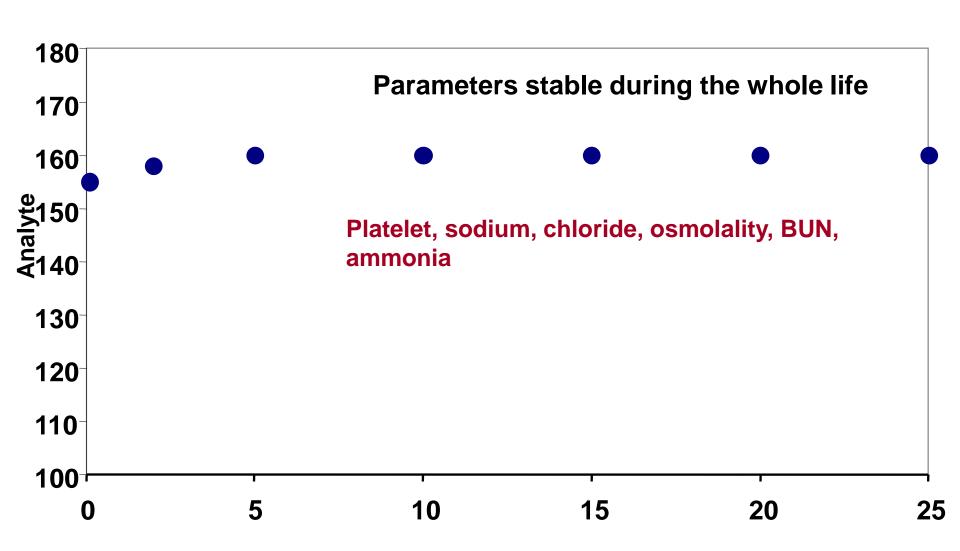
- Currently: app. 400 parameters measured routinely
- Semmelweis University: 6 million test per year
- Incidence of unnecessary test: estimated
 10 40%

Efforts to decrease numbers of unnecessary tests

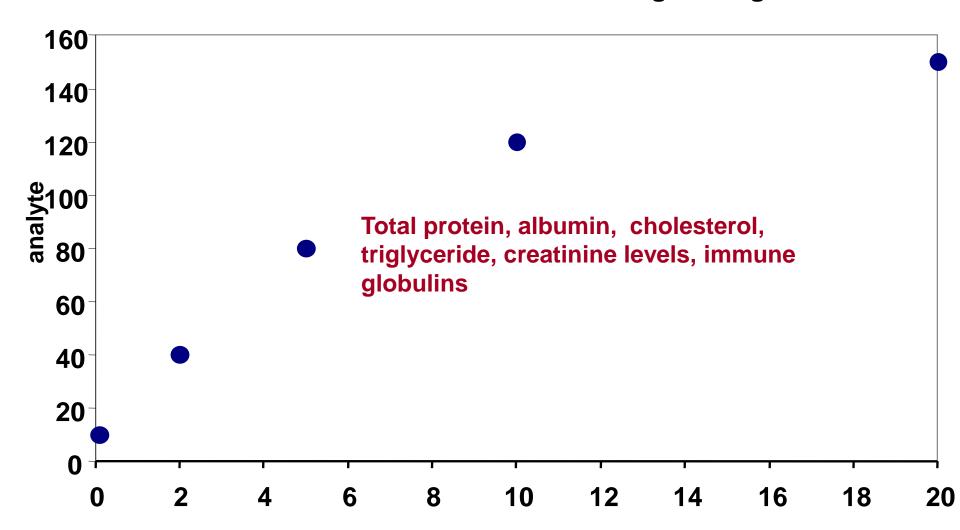
- Request linked to specific conditions
- Restricted lists
- Limitations of repeated requesting
- Education

Factors responsible for preanalytical errors - PATIENT Biological variability and environmental conditions

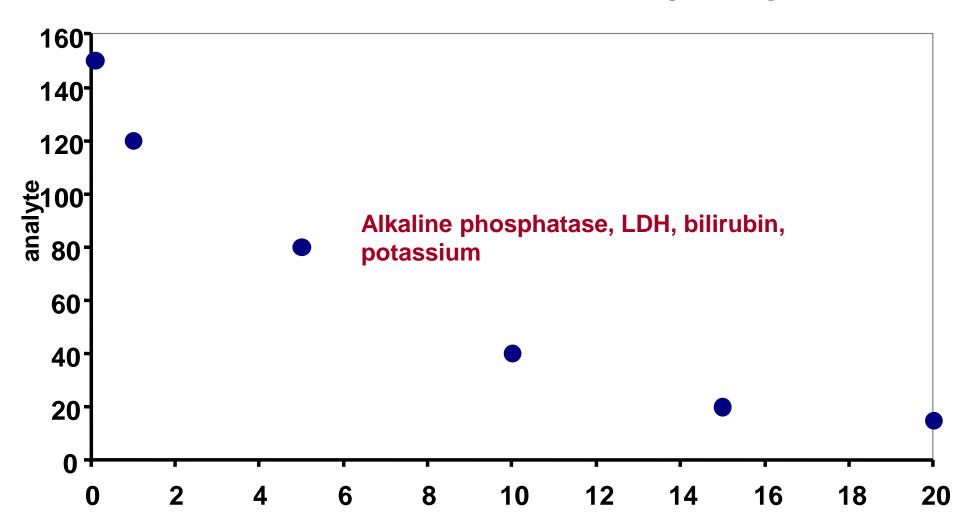
- Gender, period in fertile women
- Age
- Circadian / seasonal fluctuation
- Posture and prior exercise
- Diet



Parameters increasing with age



Parameters decreasing with age



Factors responsible for preanalytical variations - PATIENT Daily fluctuation

Maximum in the morni	ng			+ cortisol-regulated
ACTH	200%	Epinephrine	20%	cytokines & bone
Renin	140%	Hemoglobin	20%	markers
Norepinephrine	120%	Hematocrit	20%	
Prolactin	100%	Leukocyte count	20%	
Aldosterone	80%	Protein	20%	
Cortisol and related parameters	50%	Tyroxine (T4)	20%	
Testosterone	50%	Bilirubin	20%	
Maximum at noon	•			
Iron	100%	Potassium	15%	
Eosinophil granulocytes	30%			
Maximum in the evening	ng		-	
Uric acid	50%	Acid phosphatase	200%	
TSH	50%	СК	100%	

Posture

The level is increased when the lying patient stands up		
<10%	hemoglobin, leukocyte count, total calcium, ASAT, ALAT, tyroxine, IgG, IgA, albumin, total protein, cholesterol, triglycerides	
10 – 20%	hematocrit, red cell count, apolipoproteins	
>50%	epinephrine, renin	



STANDARDIZING SAMPLING

- Most common complication: bruises
- Should press swab for a few minutes
- DO NOT BEND the arm (keep extended)

Sampling error

The majority of preanalytical errors (60%) occurring with blood samples are due to sampling problems:

Insufficient quality / quantity of blood

Haemolysed or lipaemic sample	54%
Insufficient volume	21%
Inappropriate tube	13%
Clots	5%
Misidentification	1-2%
Other	4-5%

Sampling in fasting state: WHY is it important?

Adults: 12 hour fasting state is recommended:

Alkaline phosphatase

Cholesterol

Dopamine

Iron

Glucose

Uric acid

Insulin

Potassium

Cortisol

CRH-stimulation test

Inorganic phosphate

Triglyceride





IF sampling is done after a meal: lipemic serum that interferes the tests



Hemolysed samples

Cca 3.3% of clinical samples are haemolyzed		
64%	<0.5 g/l Hb	
31%	0.5 – 3 g/l Hb	
5%	> 3 g/l Hb	

About 40-70% of sample inappropriate for measurements are haemolysed.

Distribution of test ordering departments of 8440 haemolysed samples

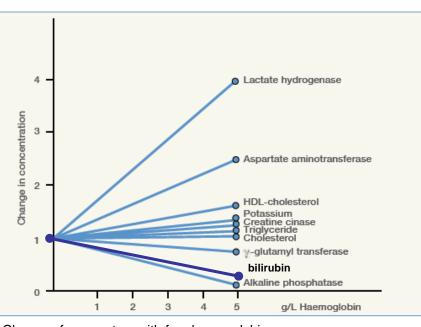
Ambulatory patients	0,1%
Dialysis	1,5%
Intensive care	5,4%
Pediatric ward	8,6%
Emergency department	8,8%

HEMOLYSED SAMPLE - DEFINITION

Free hemoglobin, upper limit	0.02 g/l (plasma) 0.05 g/l (serum)
Visible haemolysis: (when 0.5% of red cells is haemolysed)	> 0.30-0.50 g/l



HEMOLYSED SAMPLES



Change of parameters with free haemoglobin



Samples with varying degree of haemolysis

- 1. Serum levels of intracellular analytes increase
- 2. Haemoglobin interferes with photometry
- 3. Hemolysis interferes with chemical reactions

RECOGNISING THE HAEMOLYSIS

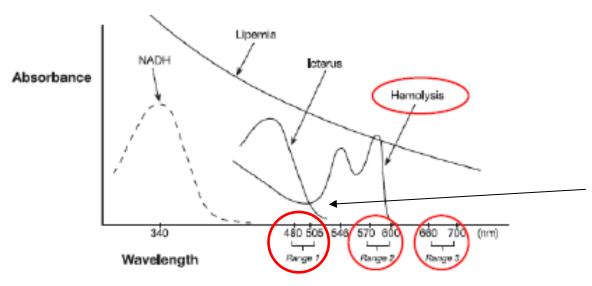
- 1. Inspection of sample (56%)
- 2. Measurement of hemolysis index (43%)
- 3. Potassium levels

Often unvisible

Interobserver variability (40%)

Jaundice may interfere

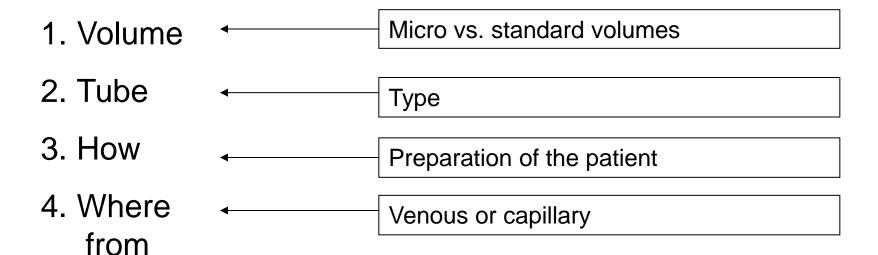
Over- or underestimated in serum or plasma



Specific absorbance spectrum should be obtained before measurements (option with clinical chemistry analysers) – haemolysis and lipemic indexes

PREVENTION OF HEMOLYSED SAMPLES: STANDARD CONDITIONS FOR SAMPLING

Planning for sampling:



VOLUME

- Volume of sample is particularly important for preterm newborns (blood volume: 80 ml/kgbw)
- Required volume is small, just a few millilitres
- Special care is required to spare sample.

TUBE TYPES



SITE of SAMPLING

- Venous sample: hand, cubital vein.
- To be avoided: infusion tubing.
- If tubes are used, this fact should be documented.
- Single tests in infants: capillary samples

STANDARDIZING SAMPLING CONDITIONS

IMPORTANT FACTORS at venous sampling

- appropriate needle size.
- strangulation: up to 1 min, 3 4 cm distance from sampling site

Increase by 6-12%	Decrease by 4%
AST, CK, bilirubin, LDH, albumin, gammGT, ALP, TP, K, Mg	glucose, inorganic phosphates, leukocytes, BUN, creatinine, chloride
HEMOLYSIS!!!	
Blood cells Protein molecules	Calcium Protein bound substance

CAPILLARY SAMPLING

Capillary sampling:

Beneficial: POCT, anxious patients, 'bad' veins

Pricking: for monitoring, single tests (e.g. blood glucose)

Incision: less pain, quicker, less traumatic. Appropriate for simultaneous tests.

Important: glucose levels are higher, potassium, total protein levels are smaller than those in serum.



CAPILLARY SAMPLING I.

- 1. First drop should be sweapt (high tissue fluid content)
- 2. Capillary should be in contact with blood drop, not with the skin.
- 3. Keep the capillary horizontally
- 4. Forced massage should be avoided (haemolysis, tissue fluid contamination)
- 5. Tube containing anticoagulants: turning several times.







CAPILLARY SAMPLING II.

Under 1 year of age: side of the plant. HEEL or FINGER are not appropriate After 1 year of age: finger III or IV, distal part.

Should be avoided: ear, thumb, index





Technical aspects of capillary sampling

Cleansing the area

Isopropil alcohol, 70%. (Ethanol should be avoided as it damages the skin at small infant)

DO NOT blow the area.

Wait until it dries up (risk of haemolysis)

Improving the blood supply

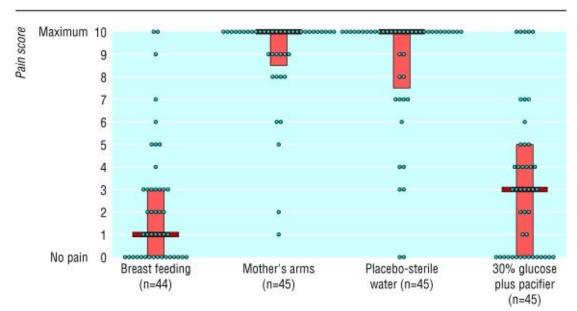
Prewarming the area with 38 °C water. DO NOT rub.

SAMPLING CONDITIONS

Preparation, calming (anxiety is a barrier for sampling)

Before 1 year

Calming the infant: oral glucose / saccharose before sampling [Metaanalysis of 14 trials: pain decreases by 20%; 12 sec shorter crying] Breast feeding has comparable efficacy



STANDARDISING THE SAMPLING CONDITIONS

Between 1 and 3 years of age

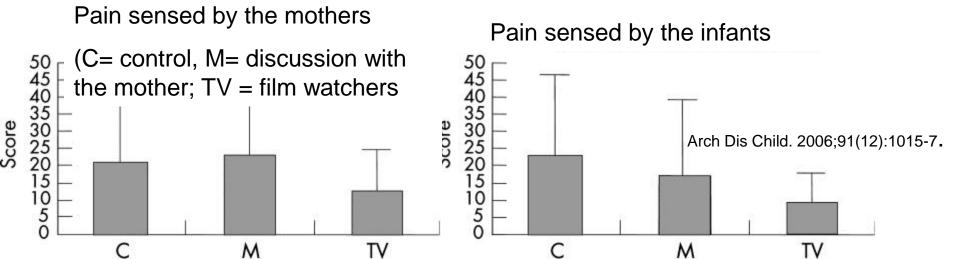
If the parent is appropriate, (s)he should keep the infant.

The phlebotomist should act quickly and reassuringly.

Communication is hard.

After 4 years of age

The role of communication increases. Details should be explained (presentation of the needle, explanation of sampling equipment etc.) Film [TV] watching during waiting



SAMPLING CONDITIONS

Order of tubes:

Venous sampling // until mark

IMPORTANT: avoid tubes over the expiry data

- Blood culture
- Citrated tube for hemostasis tests
- Native tube for serum
- Heparinated blood (plasma)
- EDTA blood for CBC
- NaF tube for blood glucose tests
- Others

Capillary sampling // mark +/- 10%

- FDTA blood
- Tubes with other additives
- Native tube for serum

PATIENT / SAMPLE IDENTIFICATION

- 1. Label in the presence of patient / relative
- 2. Name, date, ID number
- 3. Harsh massage to be avoided (hemolysis, sample tissue contamination)
- 4. Regular turning of tubes with additive.
- 5. Label should be positioned appropriately.

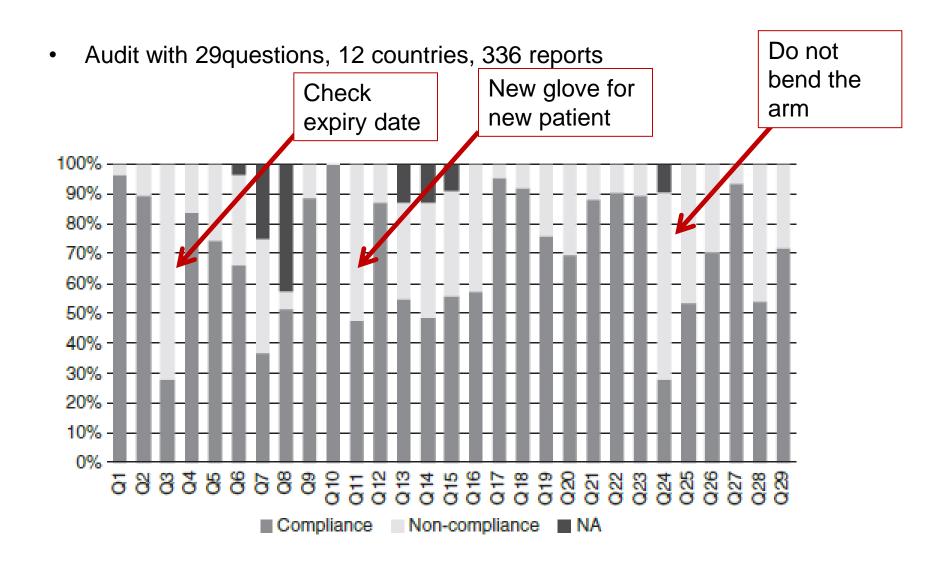




Preanalitical errors – Patient ID errors

- Study: simulated emergency department setting
- 3 volunteers: sampling, medication based on armband
- Each 3rd volunteer was given unmatched band
- 39% of nurses did not noticed this error

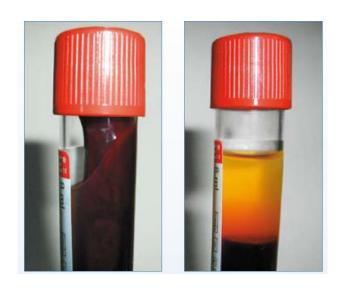
Guideline to standardize sampling processes



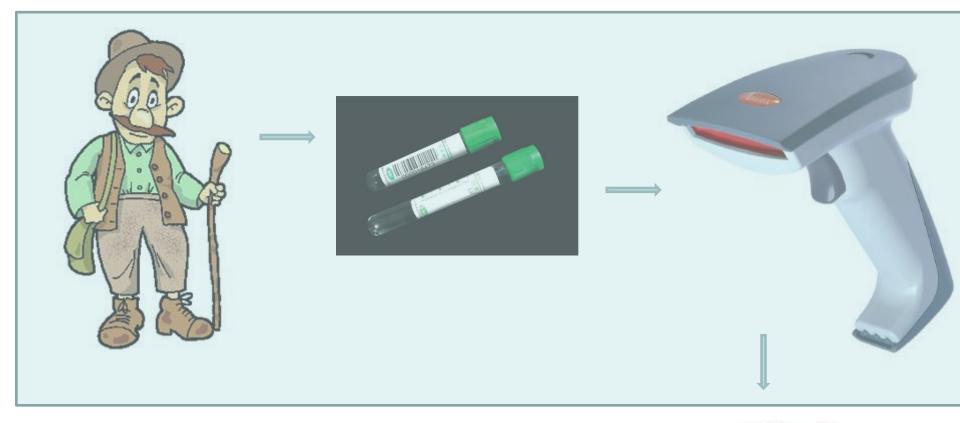
TRANSPORT AND PROCESSING

Some points to be adhered to:

- Transport in vertical position
- Do not expose to sunlight (bilirubin)
- Transferred in 2 hours to the lab
- Temperature
- The way of transport may affect some results













SUMMARY

- Several factors have an impact on quality of sample (and obtained results).
- One should remember:
 - justified request
 - identification of the patient
 - volume and sample and way of sampling
 - tube type
 - standardized setting